

OMERO

at EastBio Imaging Workshop

Dundee, January 2018

Petr Walczysko, Balaji Ramalingam

University of Dundee
The OME Consortium



Open Microscopy Environment
Centre for Gene Regulation & Expression
School of Life Sciences, University of Dundee
Dundee, Scotland, UK

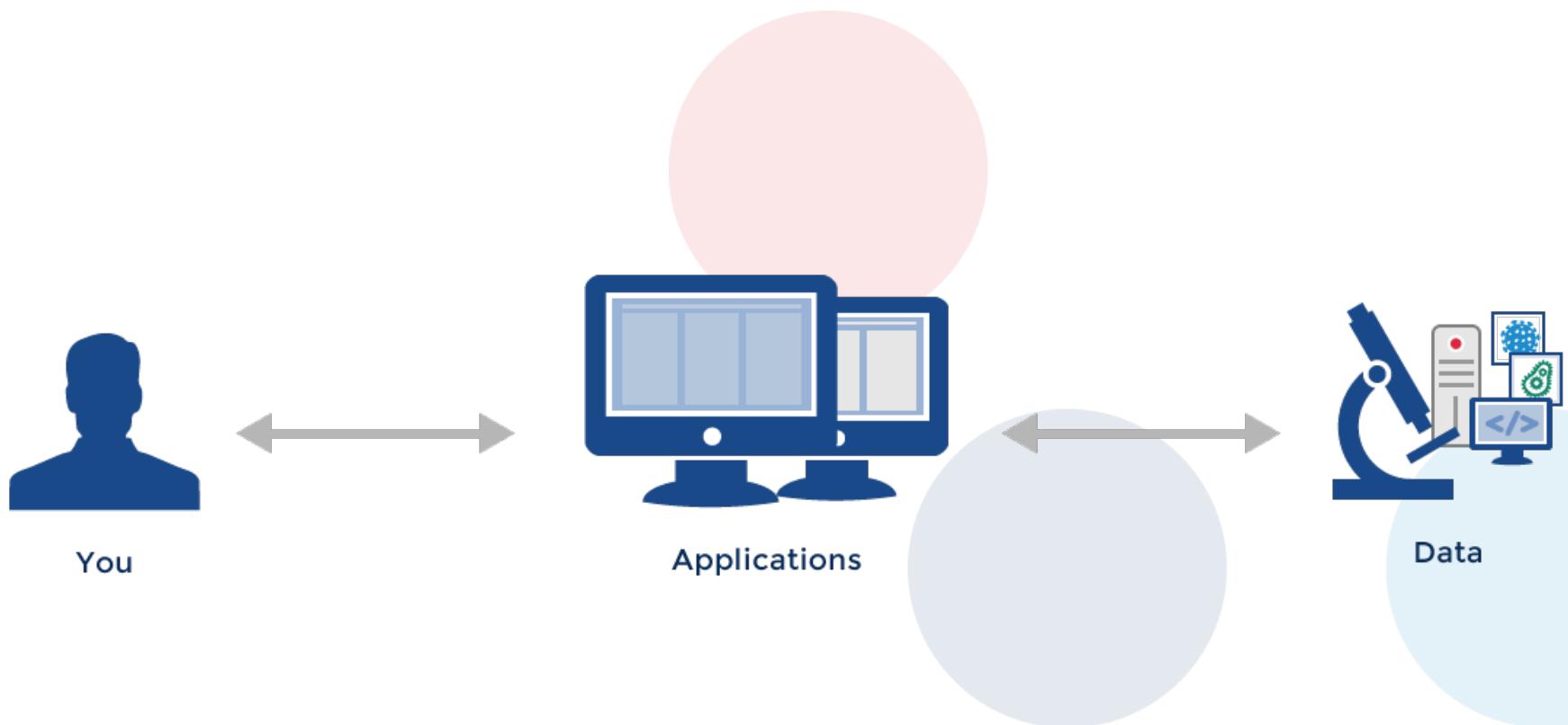
Programme of the workshop

- Short introduction to OMERO
- Viewing and managing images using OMERO plugins (OMERO.web, OMERO.iviewer)
- Data search, analysis using Fiji features (OMERO.insight plugin for Fiji, Workflow – timelapse images)
- Publishing with OMERO (OMERO.figure – Workflow - timelapse continued)

Outline

- Scientific Data paradigm
- What is OMERO
- Analyzing with OMERO
- Sharing data with OMERO
- Publishing with OMERO
- Questions

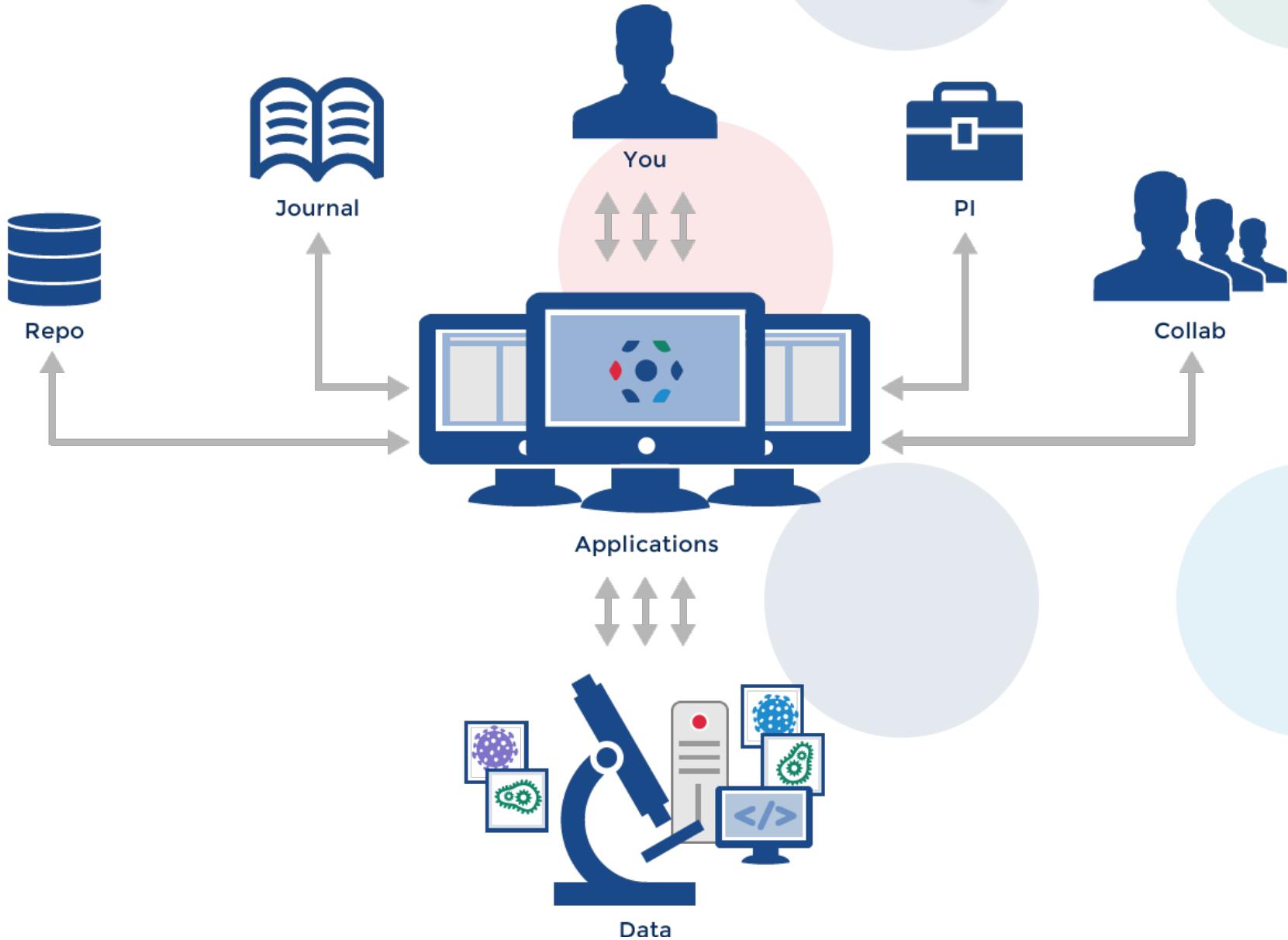
The Standard Paradigm



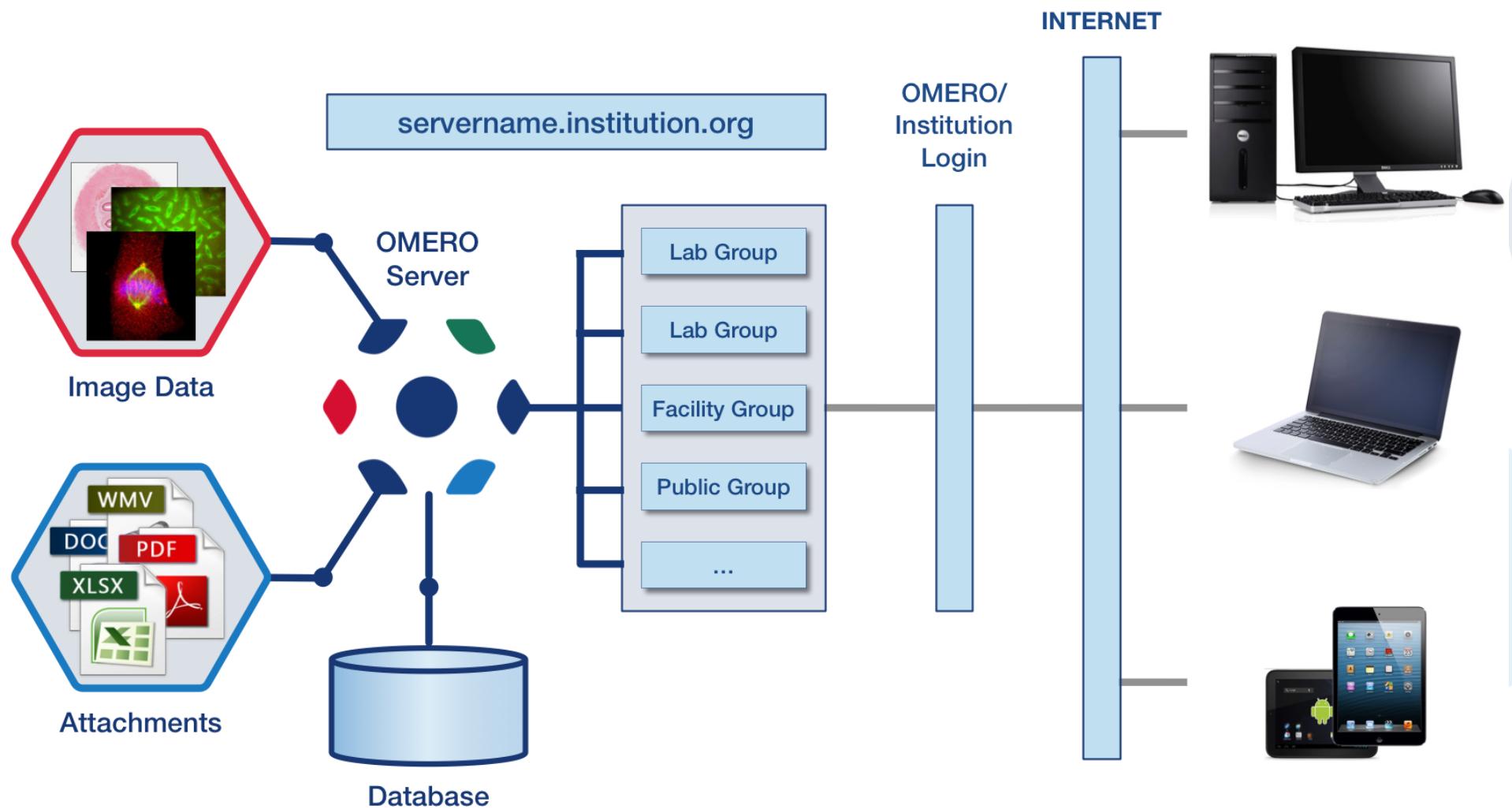
What about

- Organizing your data?
- Sharing data with coworkers and colleagues?
- Analyzing data?
- Publishing data?

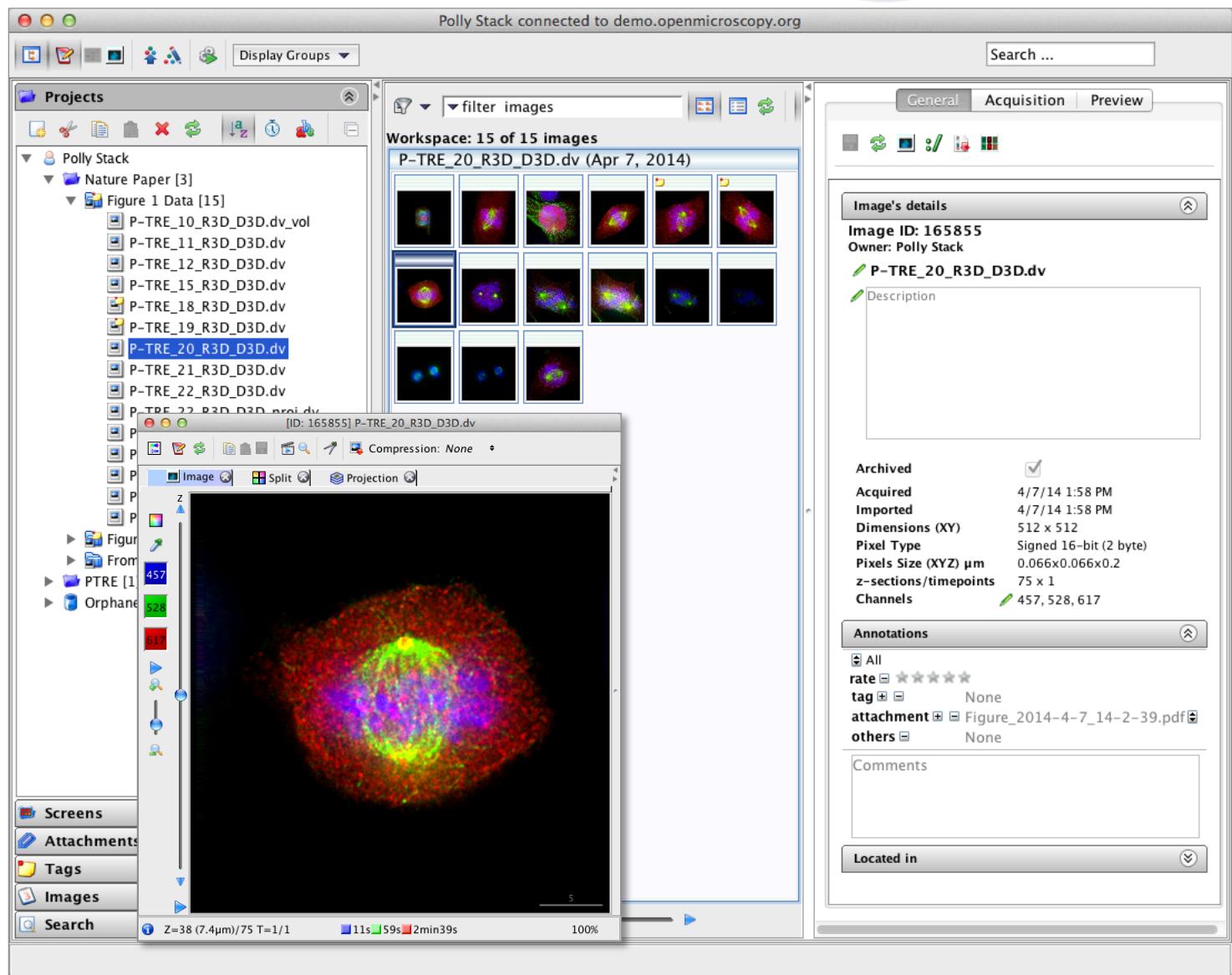
The “Scientific Data” Paradigm



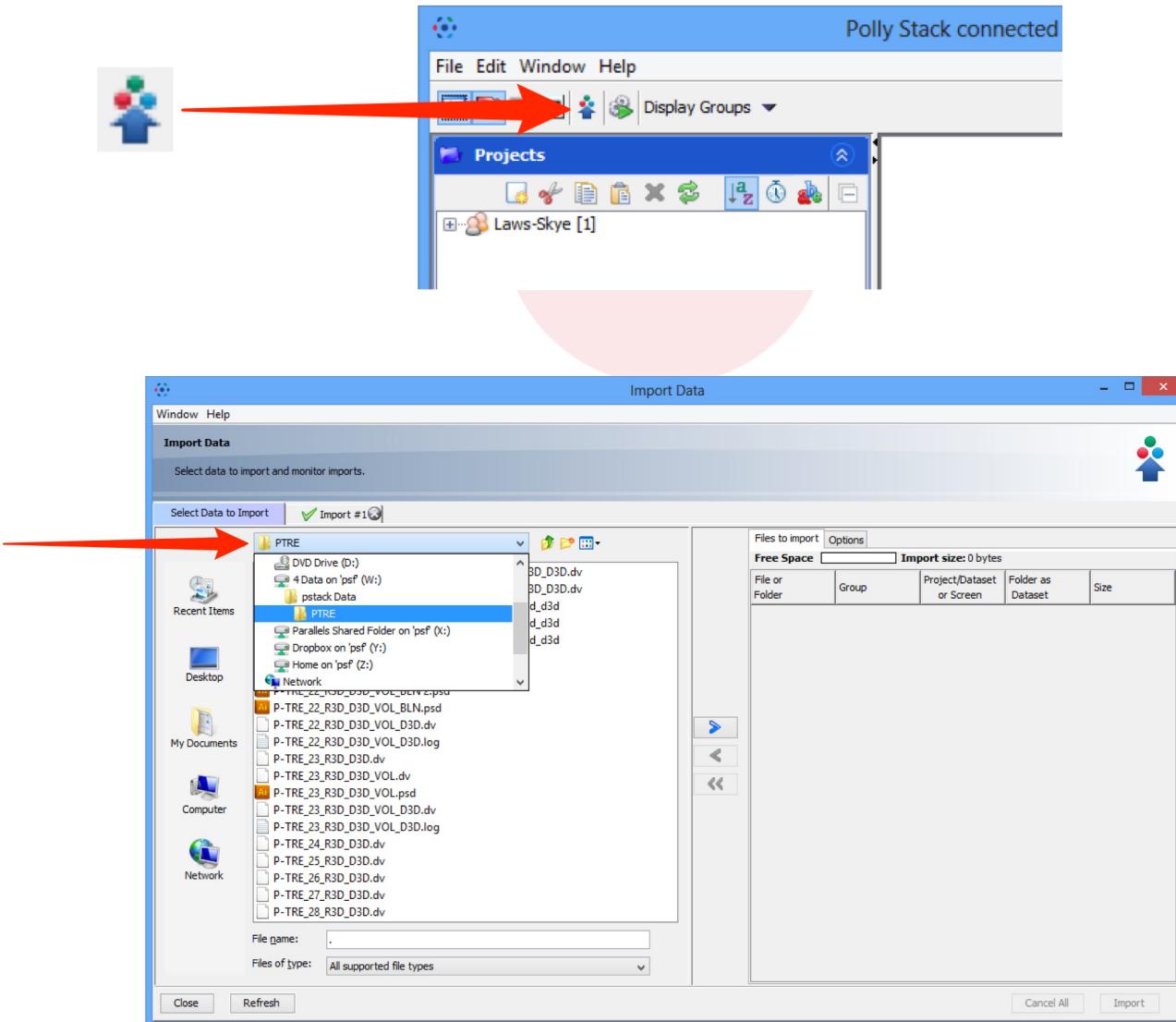
OMERO setup



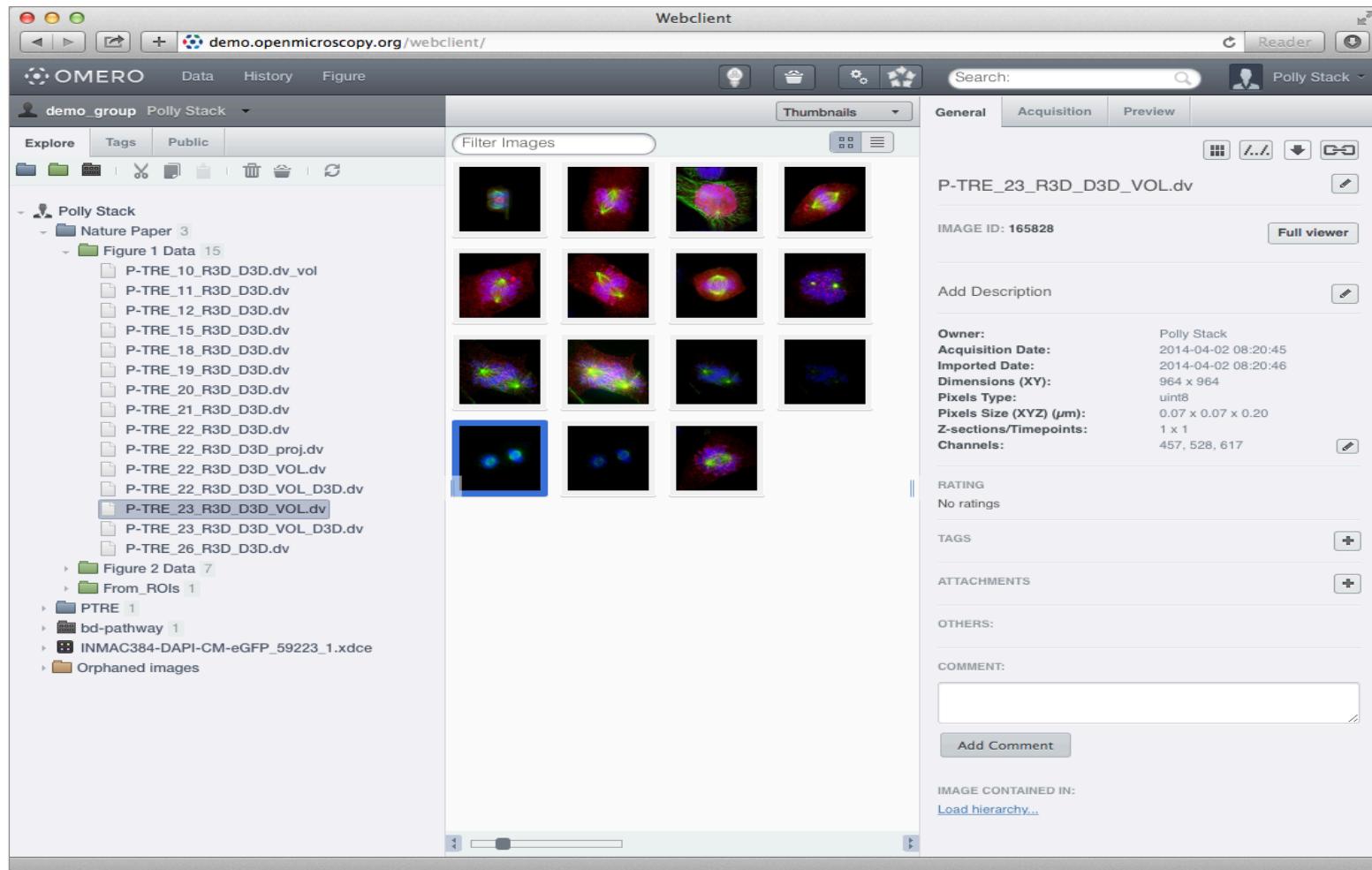
OMERO.insight: Desktop Based Application



Import Image Data into OMERO

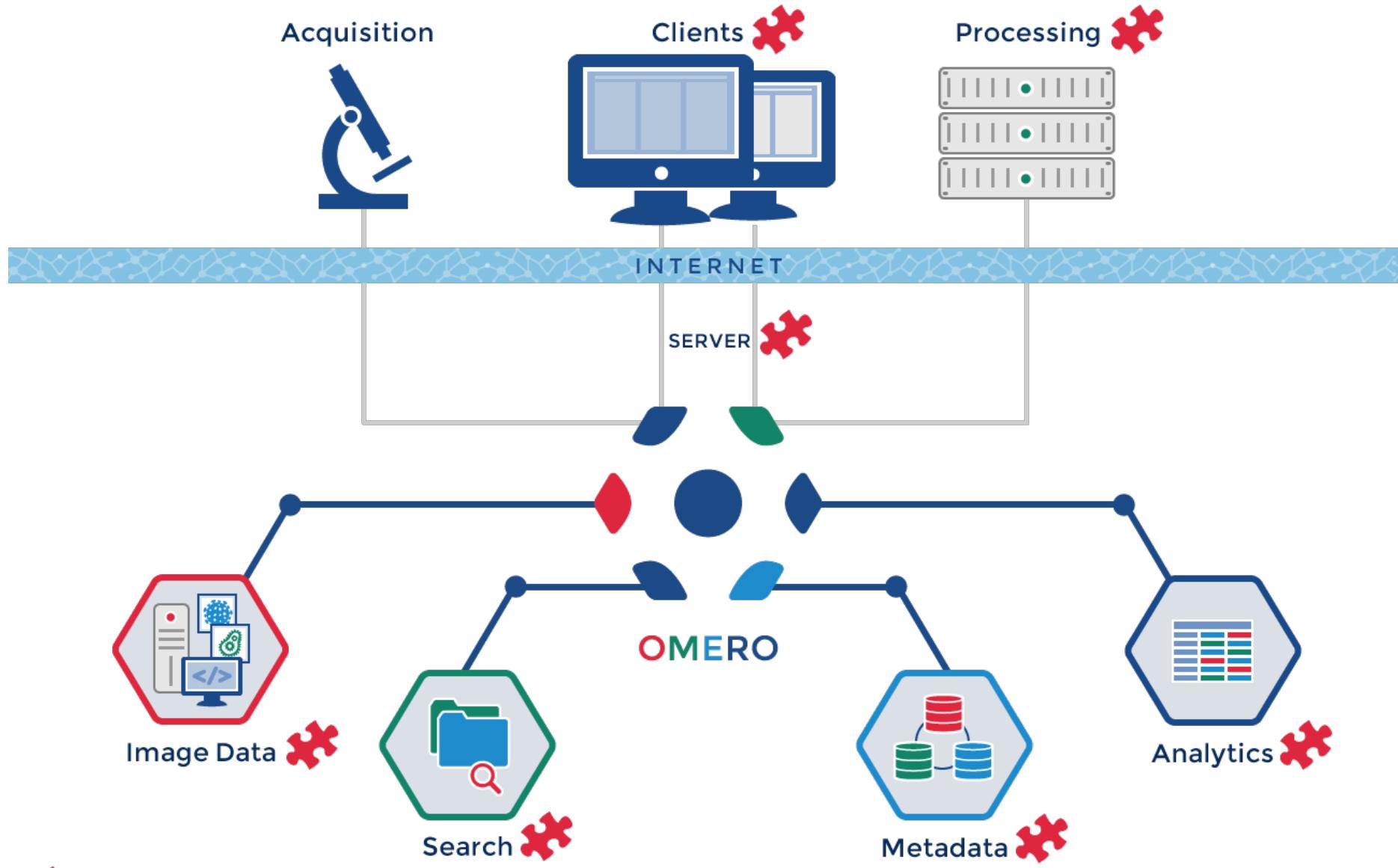


OMERO.web: Web Based Application



ANALYSIS WITH OMERO

The *Extensible* OMERO Platform



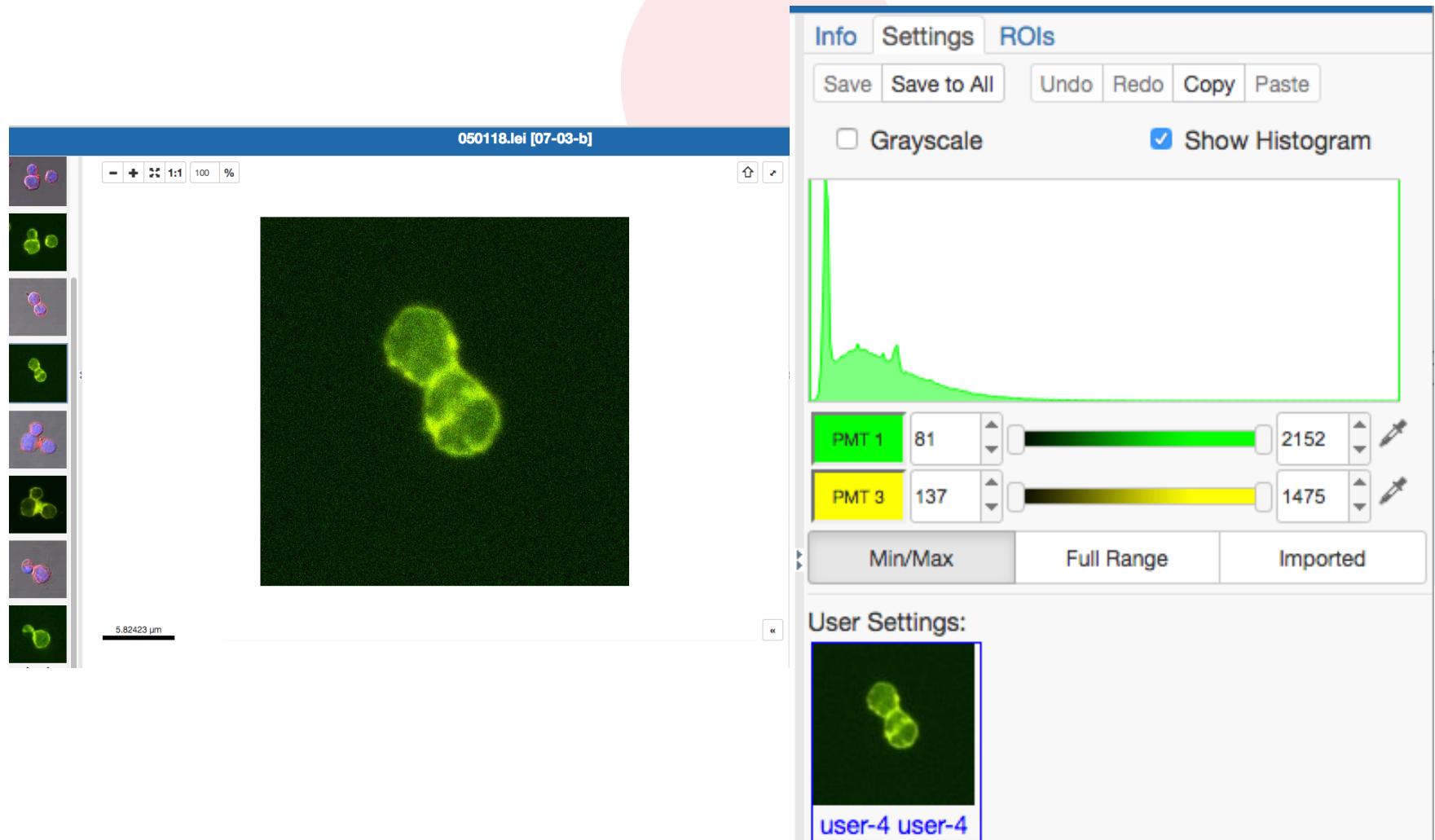
Plugins Welcome

Examples of Analysis Integration

- **OMERO.iviewer** – plugin for viewing and analysis
- **ImageJ/Fiji, Icy**– Pluggable, desktop Image processing tools (Java)
- **FPBioimage** – 3D viewer from Cambridge
- **R** – statistical analysis software
- **CellProfiler**– HCS segmentation and features (Python)
- **mTools**– Otsu, basic segmentation (Matlab)
- **ORBIT** – Image analysis, specialized on pathology images
- **WND-CHRM**-- weighted nearest neighbor machine learning (Python)
- **ThunderSTORM and PALMSiever**– Localisation SRM (ImageJ, Matlab)
- **OMERO2CV**– LSFM Multi-View Reconstruction (C++, OpenCV, ITK)
- **Columbus Acapella®**-- commercial Big Data processing...

Viewing and analyzing Images – OMERO.iviewer

- *Google for OMERO.iviewer*
- *Go to YouTube and search for OMERO.iviewer*



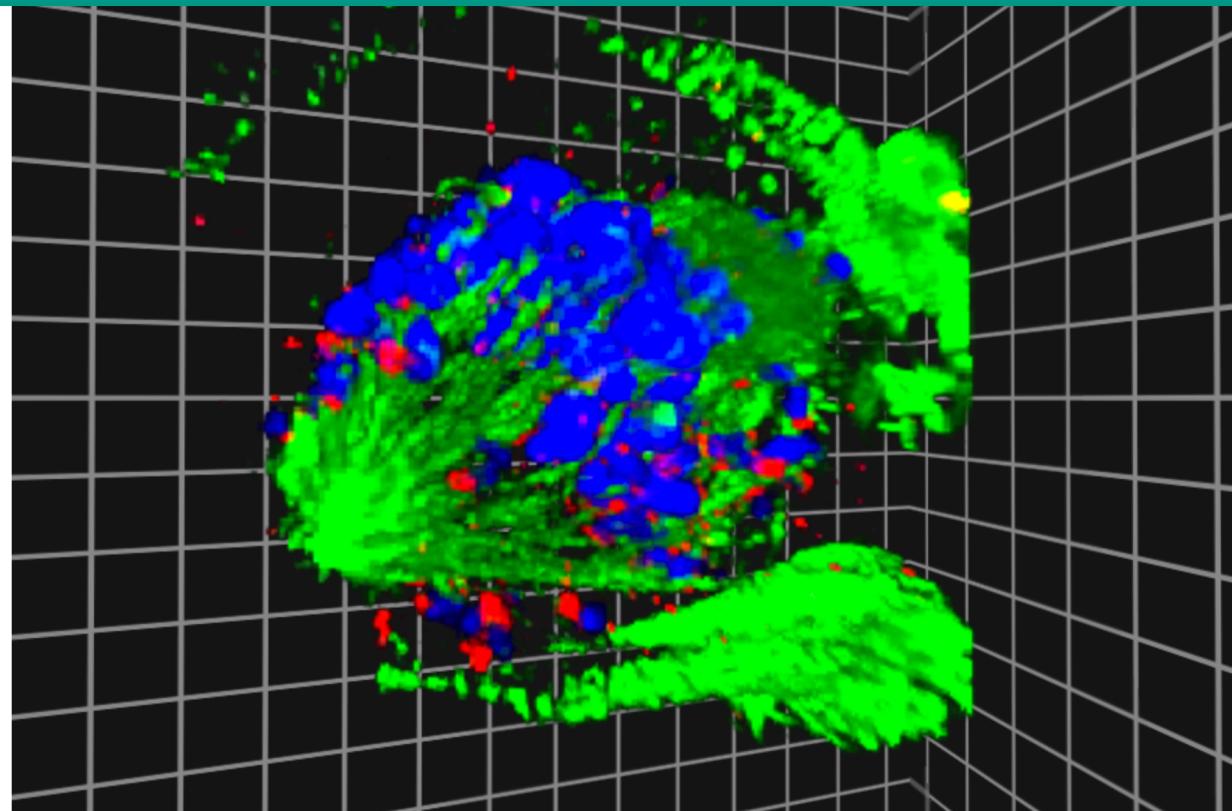
First Person Bioimage – 3D viewer from Cambridge, now in OMERO.web

© Marcus Fantham

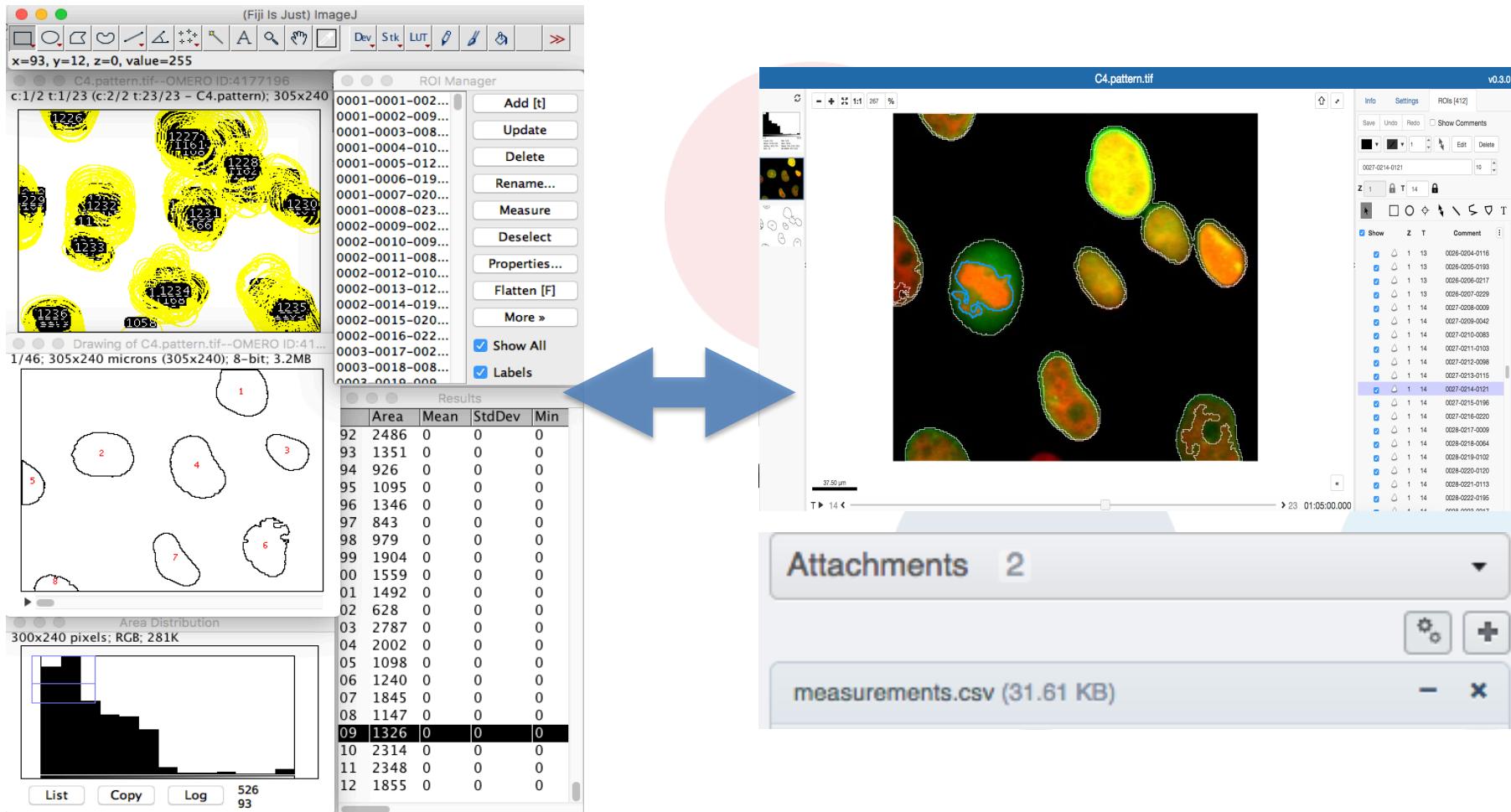
See the paper in [Nature Photonics](#)



First Person Bioimage



ImageJ and OMERO



ORBIT

- **ORBIT IMAGE ANALYSIS**
- <http://www.orbit.bio/>
- **Compatible with OMERO 5.4**

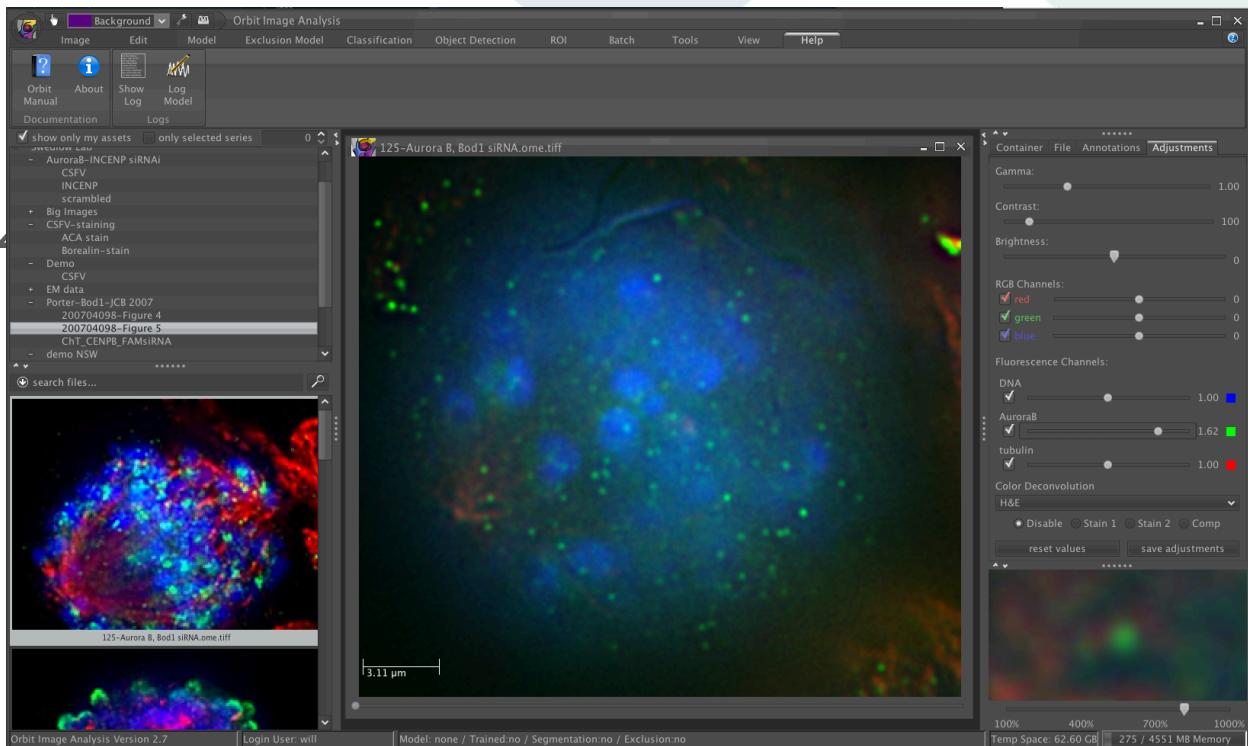
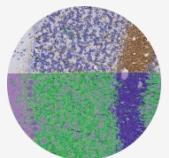


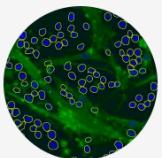
IMAGE ANALYSIS

Sophisticated image analysis features



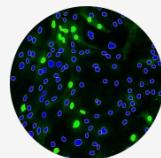
TISSUE QUANTIFICATION

Compute the ratio of different tissue classes,
e.g. percentage of collagen in a tissue.



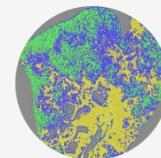
OBJECT SEGMENTATION

Segment objects like cells or nerves.



OBJECT CLASSIFICATION

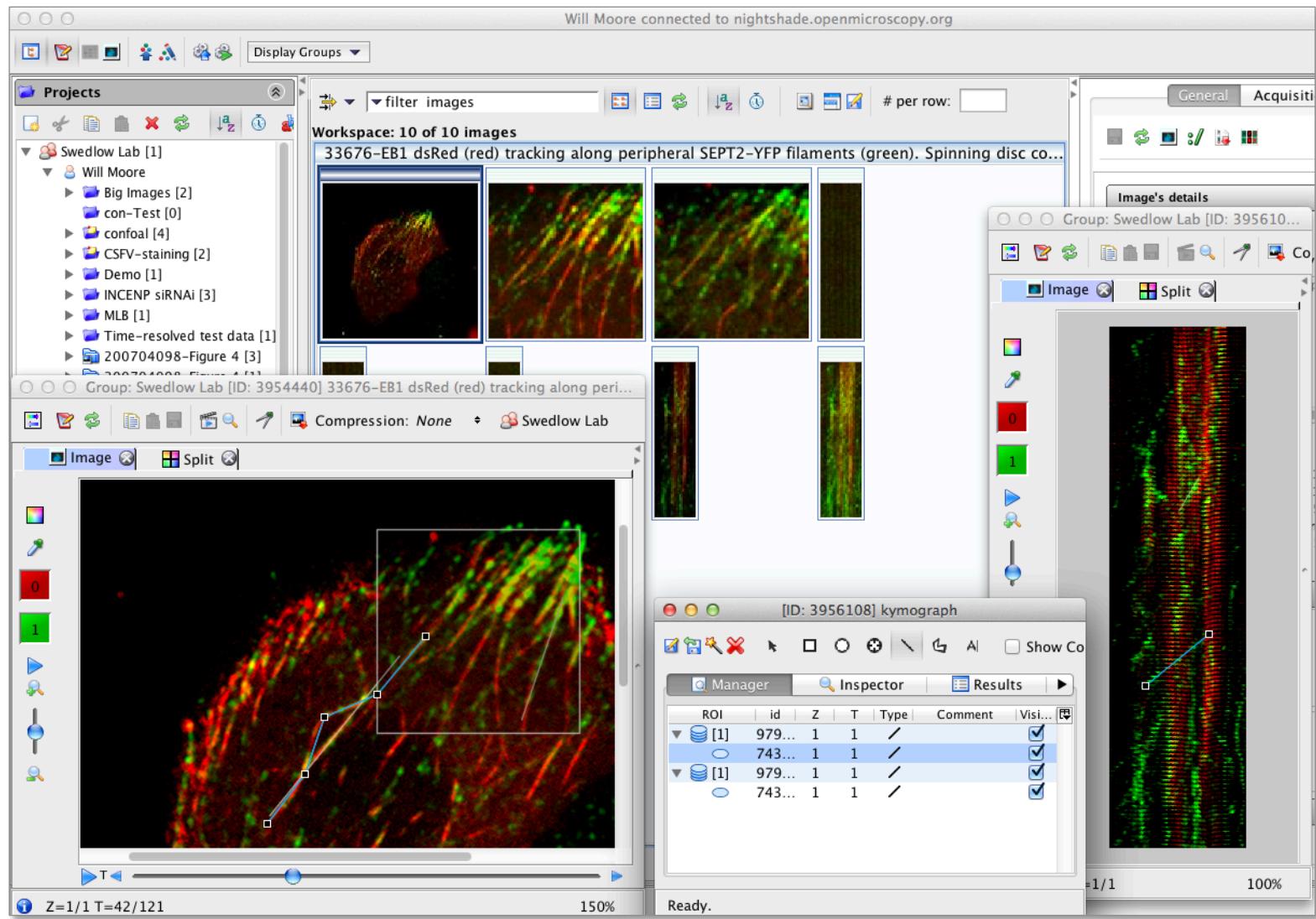
Assign classes to objects based on their
features.



ANNOTATIONS & ROI

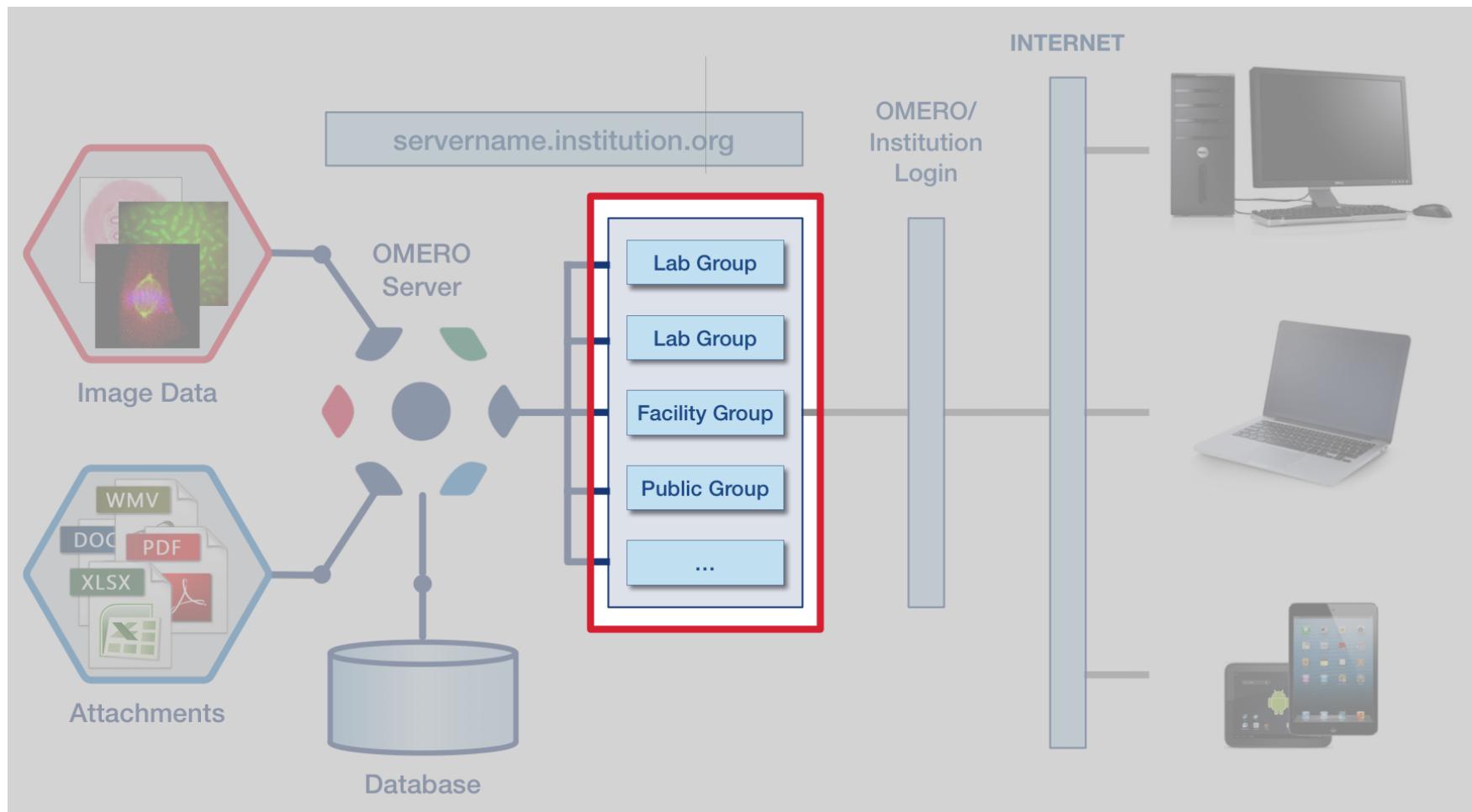
Annotations and trainable exclusion maps for
ROI definition.

OMERO/scripts: Kymographs example



SHARING DATA WITH OMERO

OMERO group and user system



Security Model



Can only read your own data



Can read but not annotate others' data



Can read and annotate others' data



Can read and edit others' data

PUBLISHING WITH OMERO

OMERO: Data Publication – images on website

The screenshot shows a web browser window with the URL https://nightshade.openmicroscopy.org/webgateway/img_detail/3933597/. The main content is a 3D reconstruction of a cell in division, visualized as a stack of Z-sections. The image is colored with red, green, and blue channels. On the left, a sidebar titled 'Viewing Options' includes buttons for 'Normal' (selected), 'Max Intensity', and 'Split Channel'; dropdowns for 'Quality' (set to 'Normal') and 'Zoom (%)' (set to 100); and checkboxes for 'Line Plot' and 'Color'. Below these are sections for 'Rendering Details' (Channels - Edit showing 457, 528, 617, with 'Color' checked), 'Current Image' (Z: 43/85 | T: 1/1), 'Image Information', and 'Image Link'. A status bar at the bottom shows 'ROI Count: 0'. To the right of the image, there is descriptive text about the cell division process and a smaller thumbnail image.

Centre for
Gene Regulation & Expression

UNIVERSITY OF DUNDEE

Home News Events Features Research Funding Impact Staff Resources Publications Contact

Jason Swedlow

P-TRE_10_R3D_D3D.dv

Position: Professor of Quantitative Cell Biology
University of Dundee, Dundee

https://nightshade.openmicroscopy.org/webgateway/img_detail/3933597/

Viewing Options

Normal Max Intensity Split Channel

Quality Normal Zoom (%) Line Plot

Rendering Details

Channels - Edit Color

Current Image Z: 43/85 | T: 1/1

Image Information Image Link

ROI Count: 0

Z-sections

Timepoints

of new daughter cells. Proper chromosome separation, and ends of microtubules. Our group studies the movement of chromosomes to microtubule attachment sites at a special time of cell division, especially in living cells and using live cell imaging techniques. We use tools to discover a new protein, Bod1, that regulates the activity of Aurora B protein kinase. We also study the centromere and kinetochore of the mitotic spindle.

In addition, we, long with our collaborators, formed the Open Microscopy Environment (OME) to develop data management software for imaging data. OME is available from the OME's web site.

View the image in OMERO by clicking on the thumbnail to view and manipulate the image in OMERO.

INCENP (red) localization in a dividing cell, also stained for microtubules (green) and DNA (blue). Click on the thumbnail to view and manipulate the image in OMERO.

© 2007-2013 Glencoe Software Inc. All rights reserved.

OMERO: Data Publication – raw data

OPEN
BIOLOGY

Advanced

Home

Content

Information for

About us

Sign up

Submit



The Ndc80 complex targets Bod1 to human mitotic kinetochores

Katharina Schleicher, Michael Porter, Sara ten Have, Ramasubramanian Sundaramoorthy, Iain M. Porter, Jason R. Swedlow

Published 15 November 2017. DOI: 10.1098/rsob.170099

The screenshot shows the OMERO software interface. The top menu bar includes 'OMERO', 'Data', 'History', 'Help', 'Figure', and 'Tag Search'. The user is logged in as 'Schleicher et al, Open Biology' under 'Katharina X Schleicher'. The left sidebar displays a file tree with a folder 'Schleicher_et al_figure2_b_d 1' containing sub-folders 'B56a ab 27' and 'B56a CTRLsi'. The 'B56a ab 27' folder contains numerous files listed as '..._B56a_B56si_01_01_R3D_D3D.dv' through '..._B56a_B56si_01_10_R3D_D3D.dv'. The main workspace shows a grid of 10x10 thumbnails, each representing a fluorescence image of a cell nucleus with red and blue staining patterns. A vertical scroll bar is visible on the right side of the workspace.

OMERO

Data Publication: Image Data Resource



Image Data Resource

Welcome to the Image Data Resource (IDR). This online, public data repository seeks to store, integrate and serve image datasets from published scientific studies.

[Take a look at the data](#)

OMERO

Data Publication: Image Data Resource

OMERO Data Publication: Image Data Resource

The screenshot illustrates the OMERO interface for managing and publishing microscopy data. The top navigation bar includes 'Data', 'History', and 'Help'.

Left Panel: Shows a tree view of 'Demo data' under 'idr0015-UNKNOWN-taraoceans/screenA'. A large grid of thumbnail images is displayed, representing a 5x3 grid of 15 individual images. Below the grid, a URL is shown: [idr-demo.openmicroscopy.org/tara/webclient/img_detail/43080/?c=1|0:255\\$FF0000,2|0:244\\$00FF00,3|0:255\\$877887,4|0:1](http://idr-demo.openmicroscopy.org/tara/webclient/img_detail/43080/?c=1|0:255$FF0000,2|0:244$00FF00,3|0:255$877887,4|0:1).

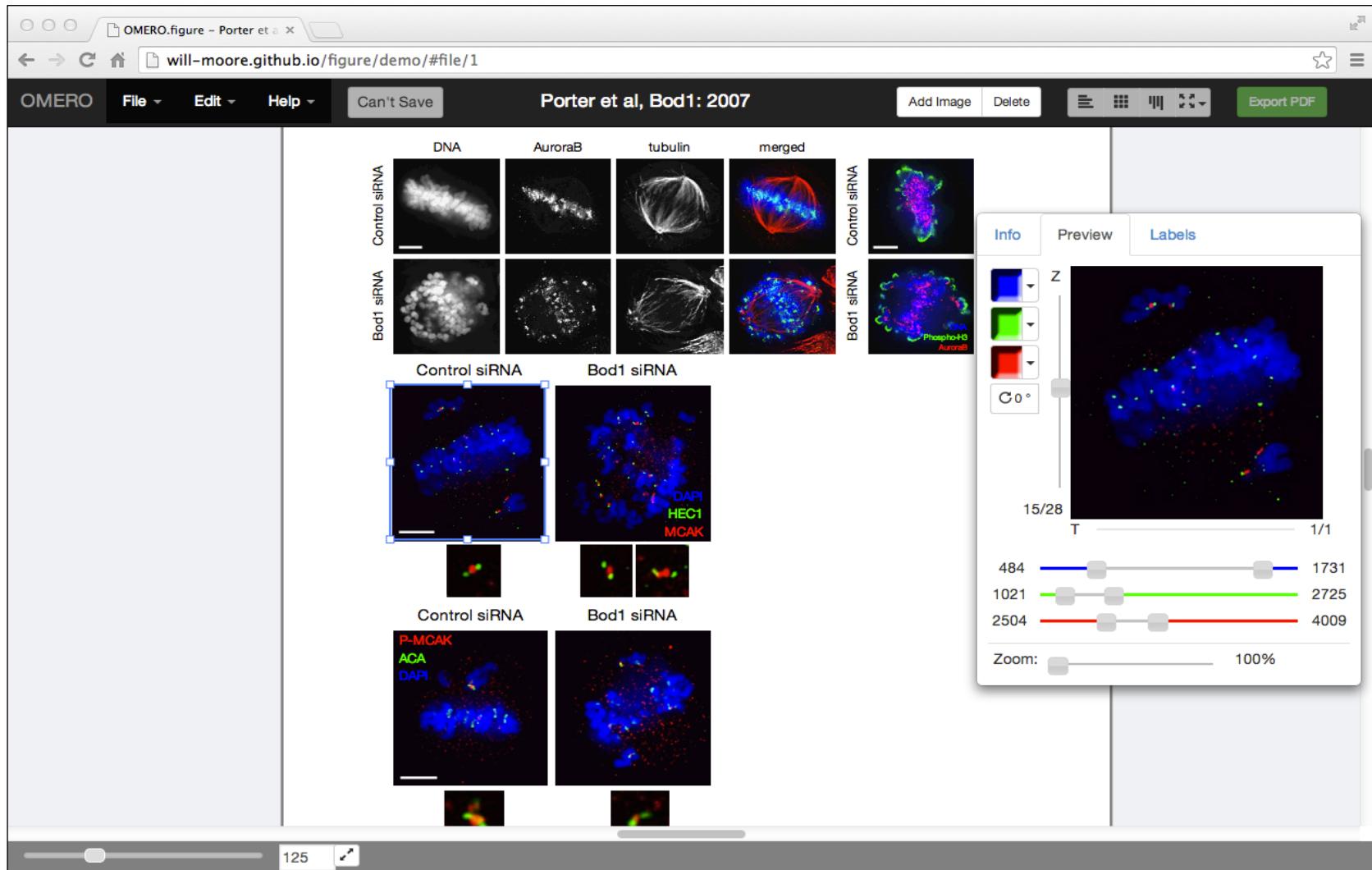
Middle Panel: Displays a detailed view of a microscopy image showing several cells. The 'Viewing Options' panel on the left allows for 'Normal' or 'Max Intensity' rendering, 'Zoom (%)' (set to 144), and 'Line Plot' selection. The 'Rendering Details' panel shows 'Channels - Edit' (selected channel 1) and 'Grayscale' checkbox. The 'Timepoints' panel at the bottom indicates Z: 11/20 | T: 1/1.

Right Panel: Shows detailed metadata for the image. The 'General' tab displays the file path: TARA_HCS1_H5_G100008302_G1000083 04-2013_12_02_21_30_23_chamber--U00--V01. It also lists 'Plate ID: 303', 'Owner: Demo User', and 'Creation Date: 2015-10-05 02:15:01'. The 'Annotations' section contains a long list of metadata fields, many of which are set to 'nan' or 'COMMENT_on_Logsheet=n/a'. A QR code is present in the annotations area.

Bottom Panel: A detailed log sheet for the 'TARA_OCEANS' cruise, dated 2009-2012. It includes sections for 'Start' and 'End' times (e.g., 2011-03-10 17:57 to 2011-09-11 01:24), coordinates (LAT DD MM.MMM, LON DDD MM.MMM), 'PUMP# DAY / NIGHT' (1 DAY), 'OPERATORS' (JP), 'DEPTH_Intended (m)', 'CABLE_Length (m)', 'Angle (deg)', 'Speed (m/s)' (SURFACE, 1.0), and a table of 'OPERATION', 'START TIME (HH:MM)', 'END TIME (HH:MM)', 'PUMP RATE (Hz)', and 'COMMENTS'. The log also notes 'Flow through GPSS (when 5μm net is not available.)' and 'PUMPING_Depth_Max (m)', 'PUMPING_Depth_Min (m)', 'PUMPING_Duration (HHMM)', and depth parameters.

OMERO.figure

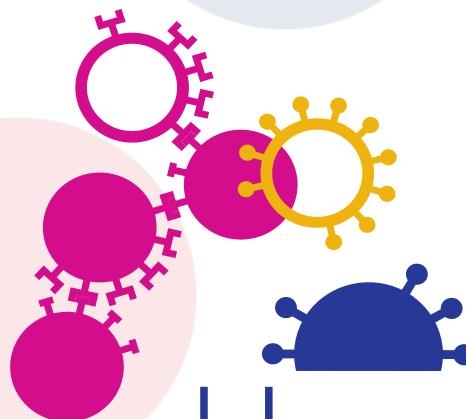
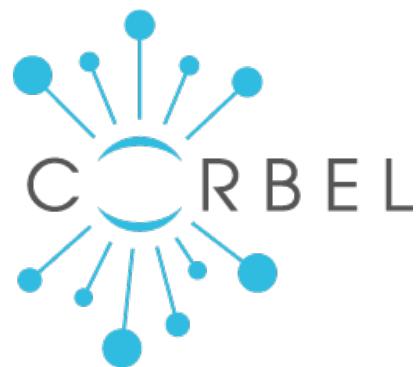
Winner of the SLS innovator of the Year



Some useful links

- OMERO Downloads:
 - <https://downloads.openmicroscopy.org/omero/>
- OMERO Help Pages:
 - <http://help.openmicroscopy.org/>
- OMERO Forums:
 - <https://www.openmicroscopy.org/community/>
- OMERO demo server:
 - <http://help.openmicroscopy.org/demo-server.html>

Thank to Funders



bbsrc

biotechnology and biological sciences
research council

