

OHSU CENTER FOR SPATIAL  
SYSTEMS BIOMEDICINE

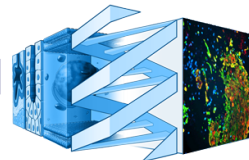


LAWRENCE BERKELEY  
NATIONAL LABORATORY:  
LIFE SCIENCES DIVISION

# Integrating OMERO into a correlative microscopy workflow: successes/struggles

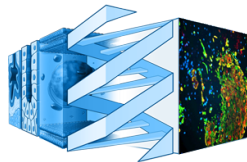
Damir Sudar and many others  
OME workshop - June 5, 2014

Oregon Health & Science University, Lawrence Berkeley National  
Laboratory, Quantitative Imaging Systems, Inc.



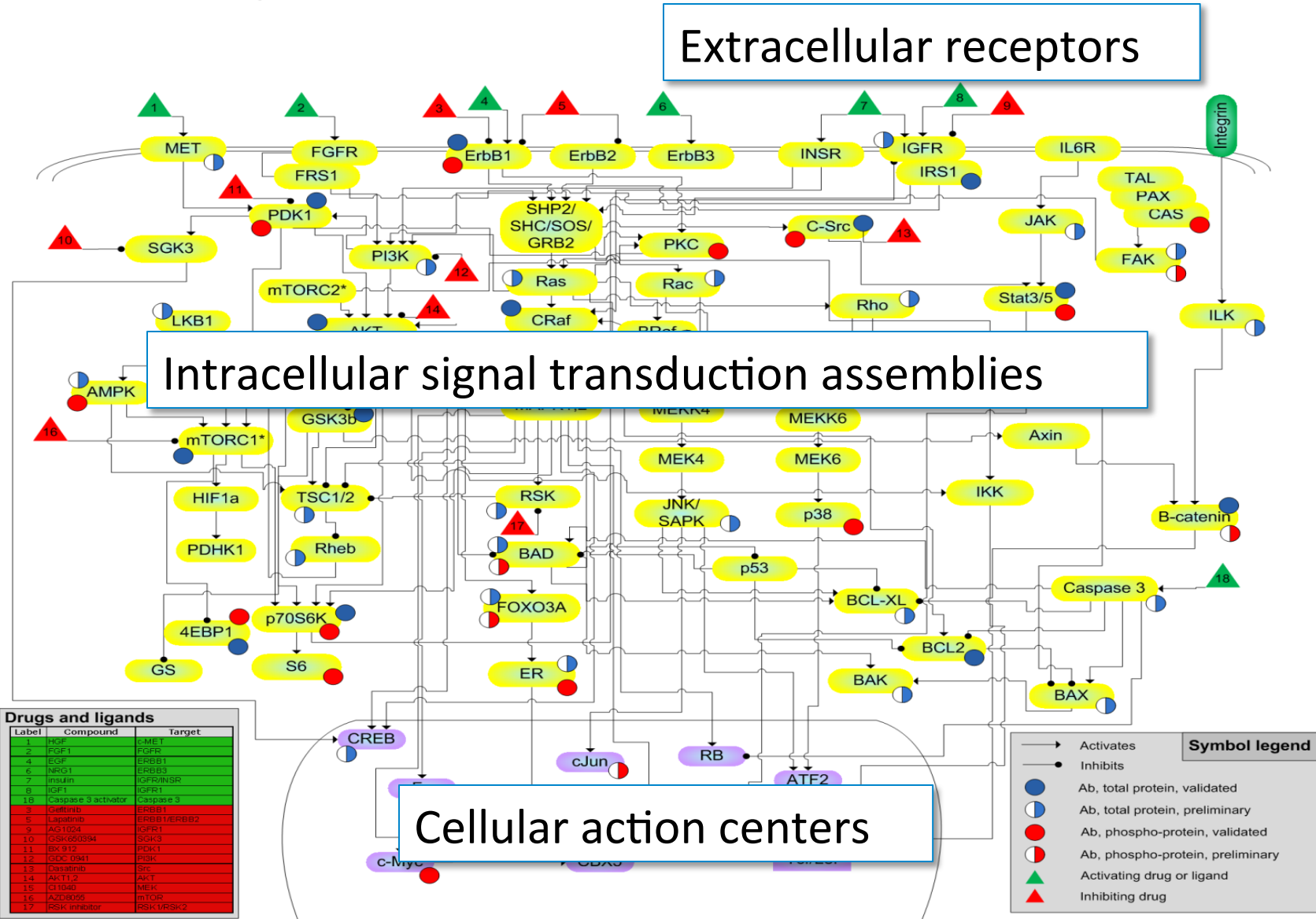
# OMERO use: Research context

- OHSU: iLEM for Multiscale Structural Epigenomics
- LBNL: Integrated Bioimaging Initiative
  - Drosophila Gene Localization Atlas
  - Next Generation Bioimaging project
  - Integrated Imaging of Microbial Community Response to External Threats
- OHSU: Various drug screens, RNAi aberration screens, and microenvironment screens



# iLEM motivation: Understanding regulatory signal transduction

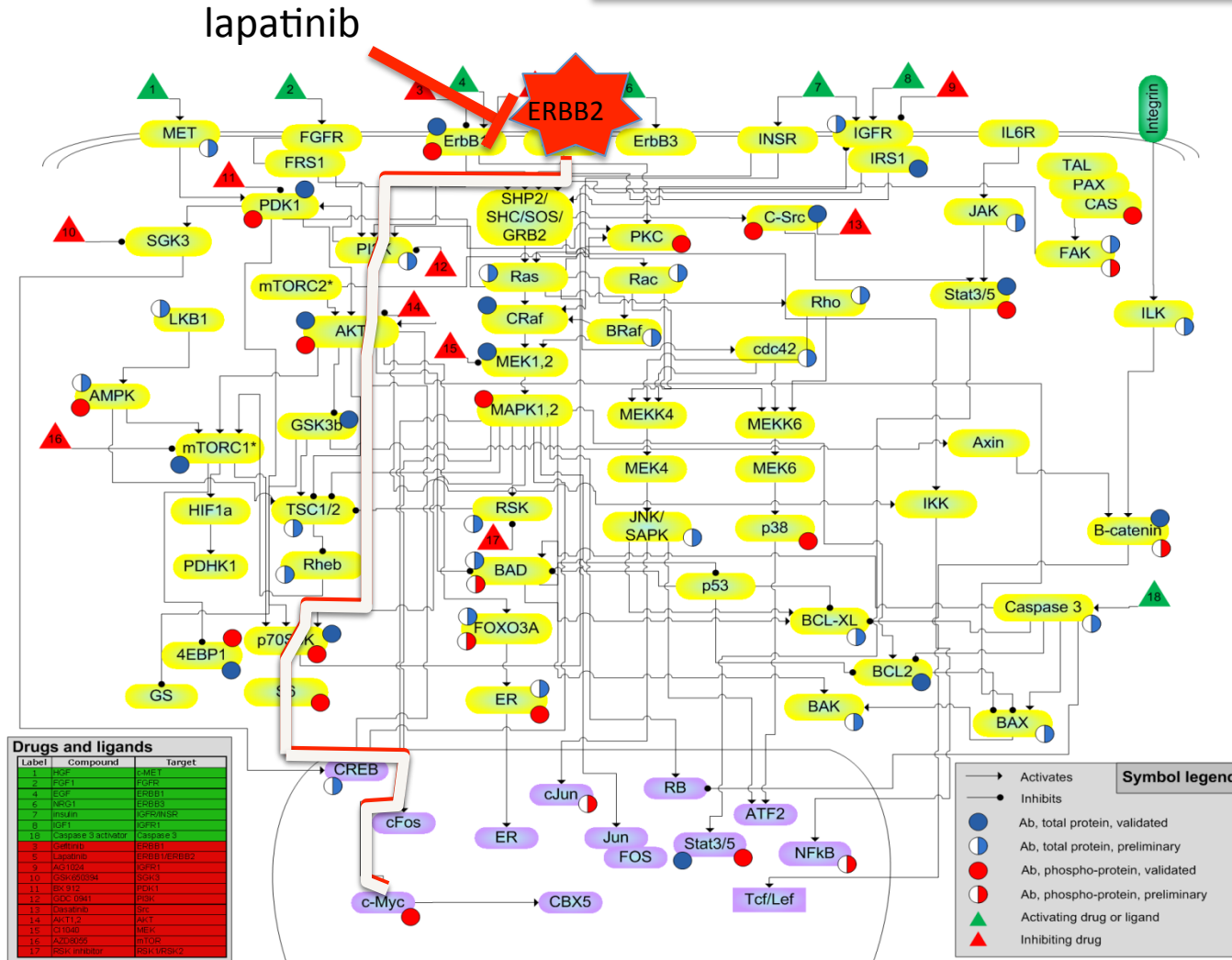
Elucidating the roles of the nano- and microenvironments



# Prototype problem

RTK signaling in breast cancer

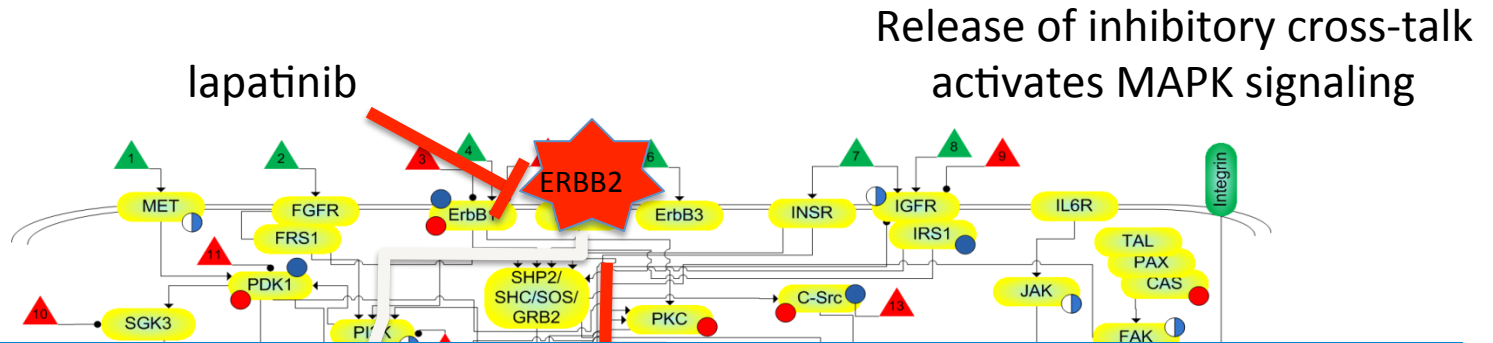
Our goal in cancer treatment is to develop therapies to correct the defect



RTK – Receptor tyrosine kinase



# Activation of bypass mechanisms can attenuate response



There are many bypass pathways

Combinatorial Rx becomes increasingly toxic unless targeting is very precise

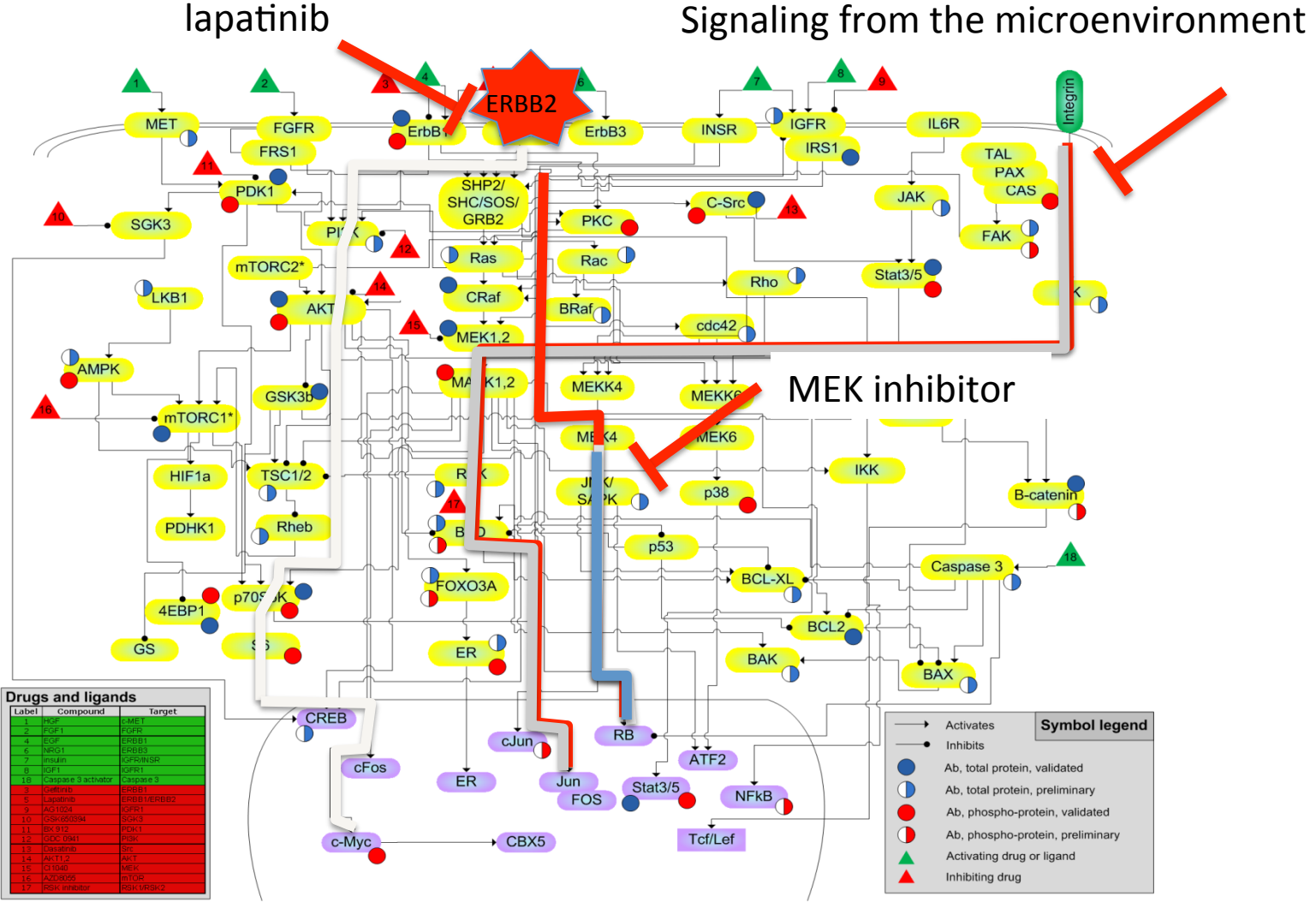
How does all of this information get integrated?

We don't know where or how it gets translated into action (proliferation, death, etc)

15	OT1608	MEK
16	AZD1775	mTOR
17	BBV-799	SH-PTEN

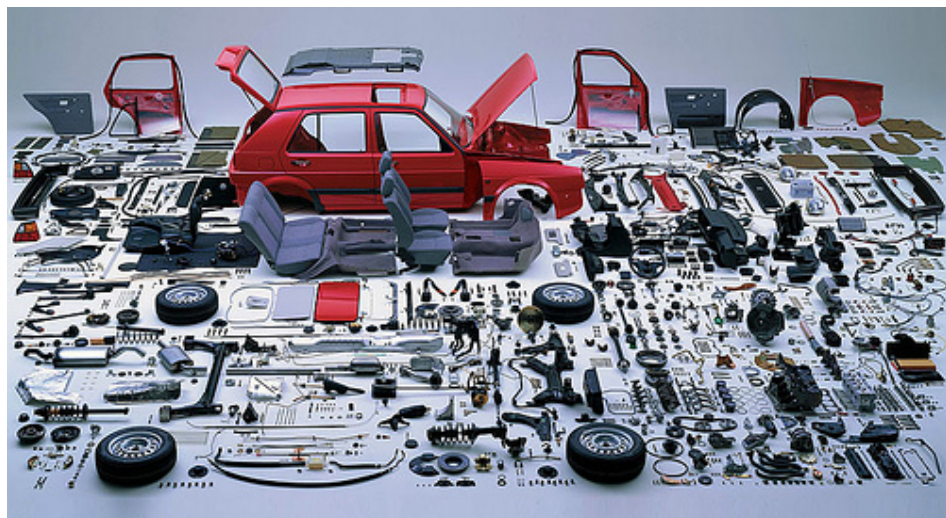
▲	Activating drug or ligand
▲	Inhibiting drug

# Signaling from the microenvironment also can bypass the inhibitor



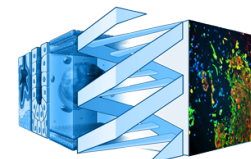
We need to know how the parts are assembled in order to fully understand function (and response to therapy)

Moving from the parts list to the assembly manual



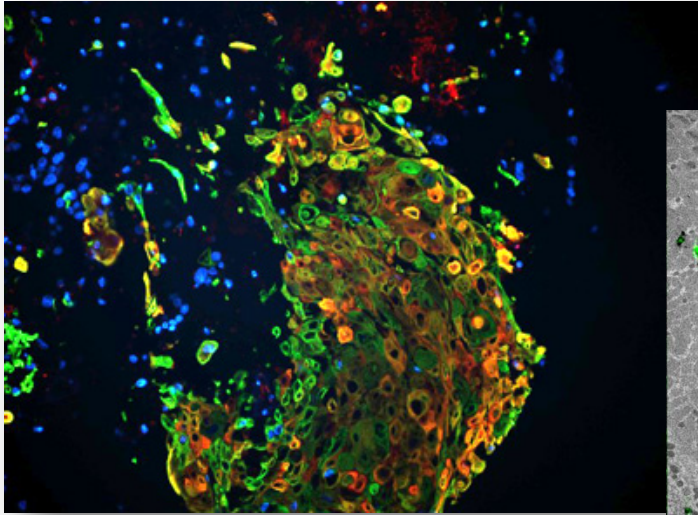
You can't understand something from looking at the parts

*You need to know how they work together*

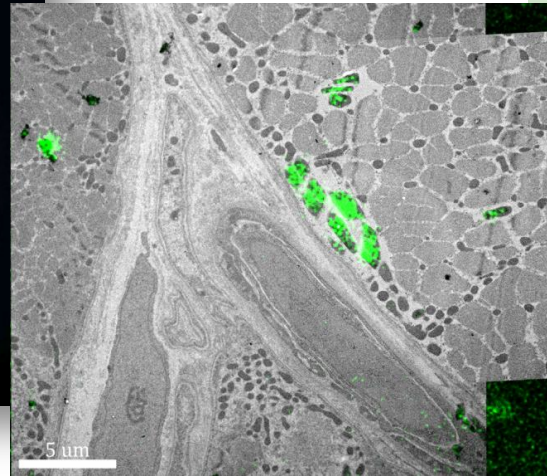


# These studies motivate the development of strategies to visualize pathways in situ

## Defining the microenvironment

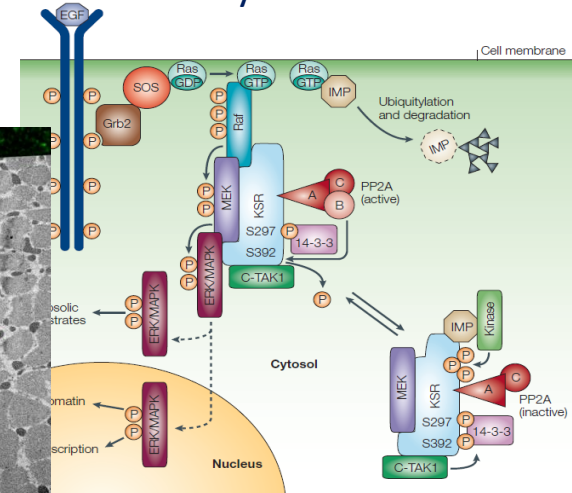


Basal breast cancer CK5/6 green, CK17 orange, CD44 red, DAPI blue



iLEM will define sub-cellular and micro-environmental context

## Pathway nanostructure



OMX

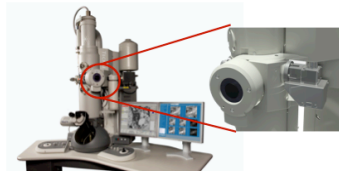


Olympus Scan^R

### Tecnai iLEM™

#### Tecnai iLEM™

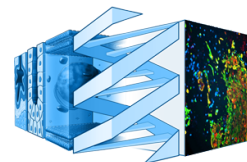
The Integrated Light and Electron Microscope; a fast, accurate, and automated solution for correlative light and electron microscopy.



FEI CorrSight



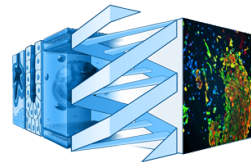
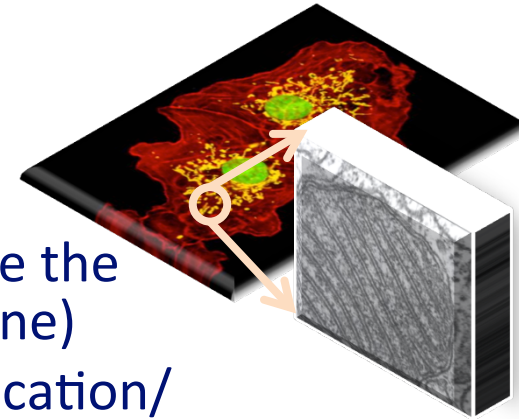
FEI Helios





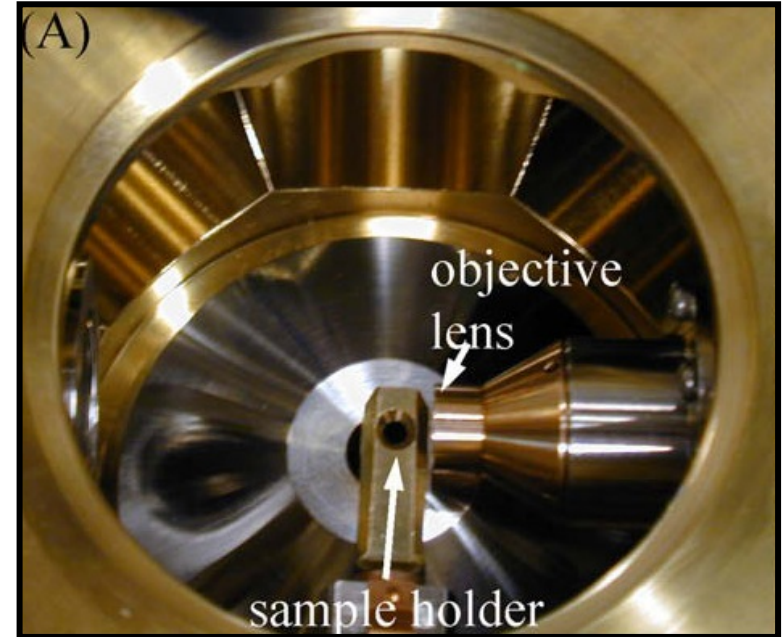
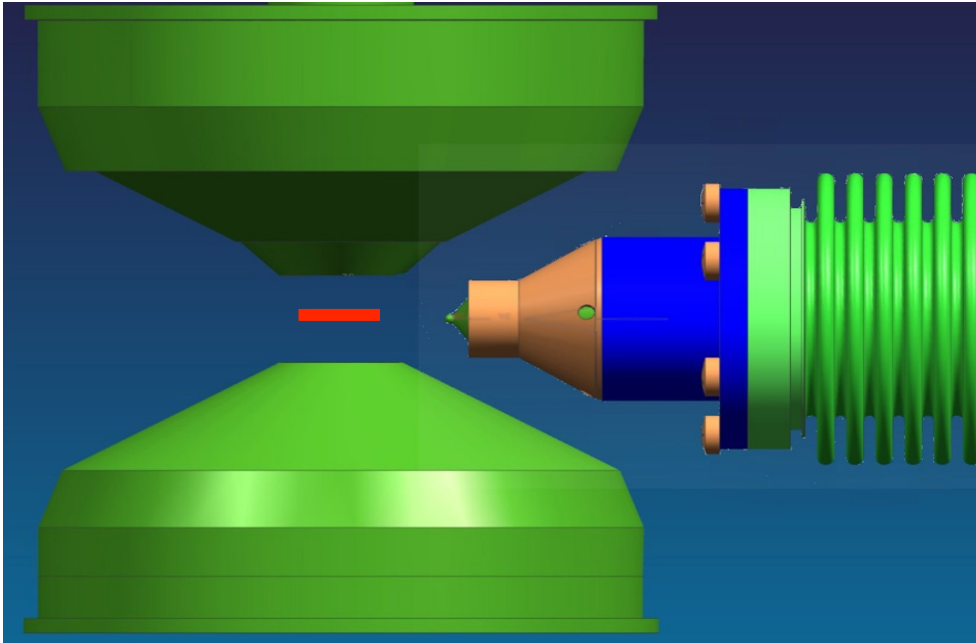
# Correlative Microscopy

- Find specific cells in a large field of view
- Find specific areas in a cell and zoom in to study these at the ultrastructural level
- How?
  - Use two different microscopes and correlate the information (two machines or combine in one)
  - Use a machine that has a very large magnification/resolution range and multimodal imaging



# Truly Integrated Light and Electron Microscopy (iLEM)

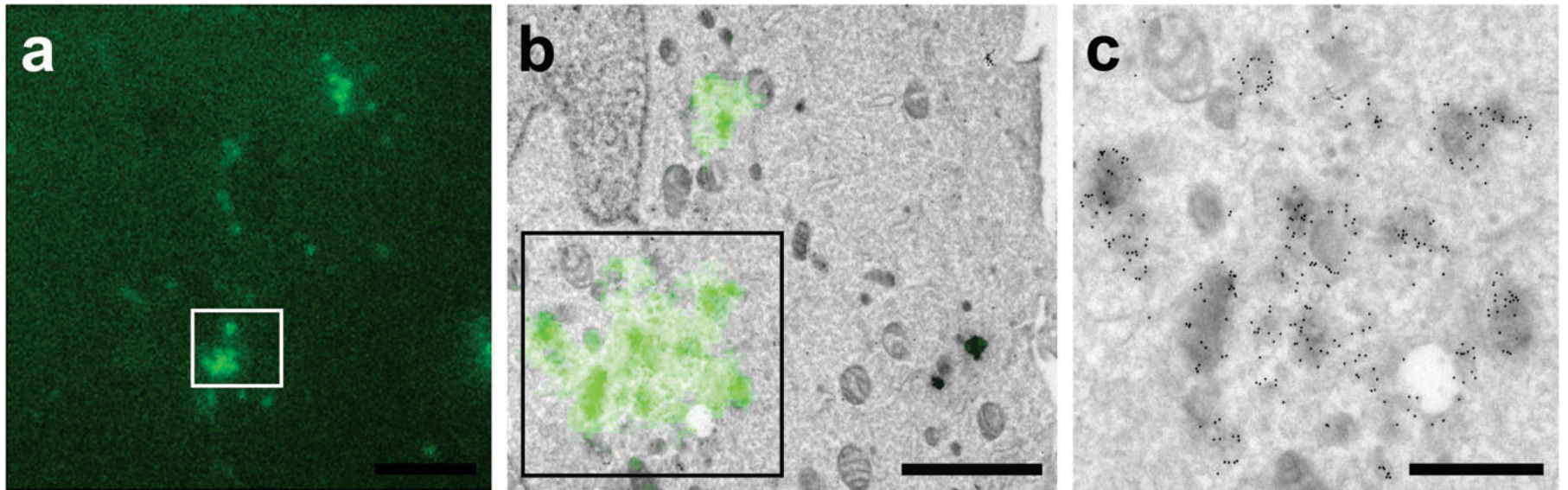
Agronskaia AV et al., J Struct Biol. 2008



## Optimal Navigation Tool for correlative microscopy

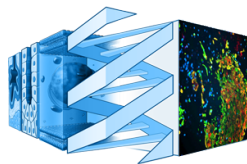
- Faster correlation between FM and EM
- Less potential contamination (cryo)
- No compromise on EM performance, decline in FM resolution

# Sample prep is critical and difficult to optimize



THP-1 cells fixed with VIS2FIX<sub>FS</sub> and IF+IG labeled for LAMP-2 (a lysosomal protein)

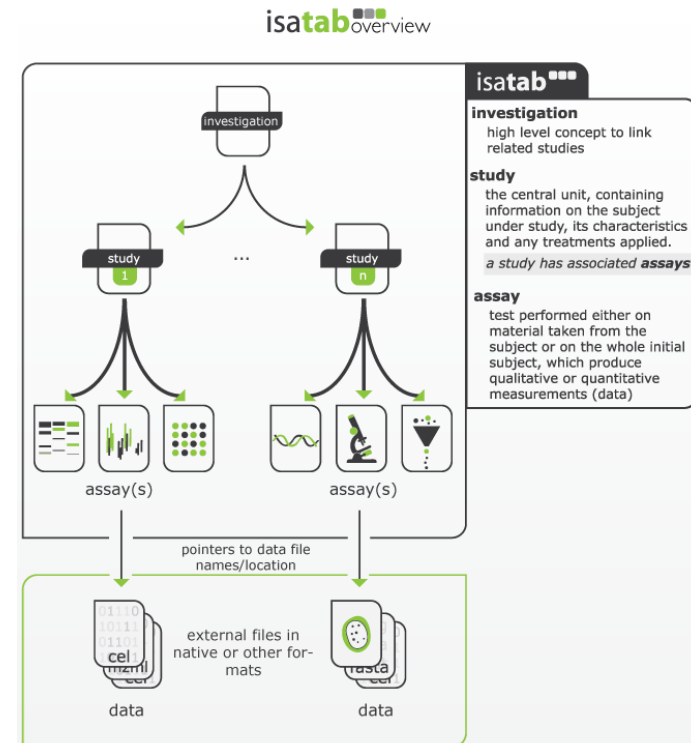
Karreman and Donselaar et al., Traffic 2011



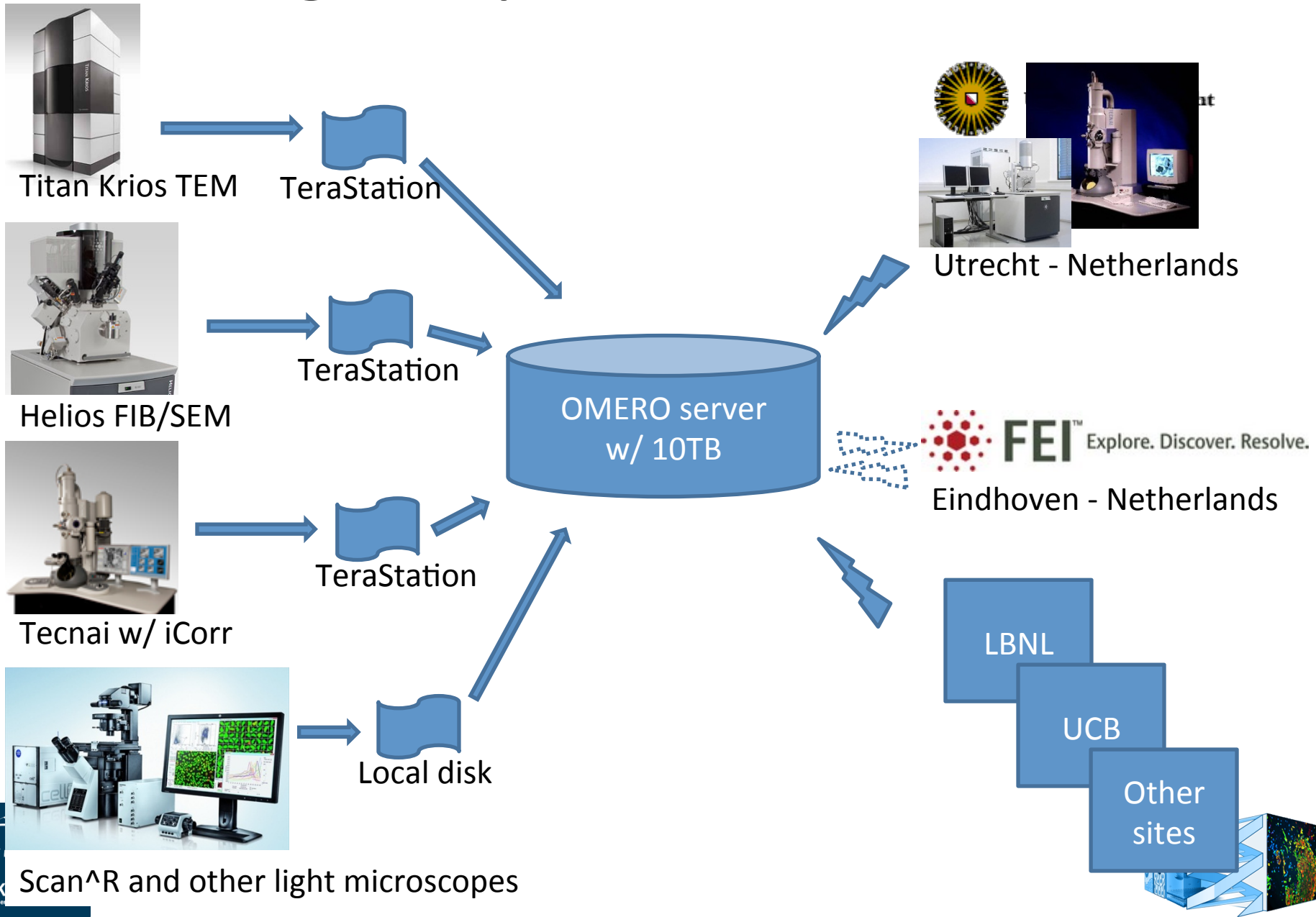
# Managing the image data, metadata, and experimental parameters

- Image data and image metadata: OMERO
- Experimental metadata: ISA-Tab & various Minimum Information formats (MIARE, MIACA, )

The screenshot shows the OMERO web interface. On the left, there is a sidebar with a tree view of folders and images, including 'conf2', 'conf3', 'confocal', 'Falp', 'Fib', 'Fim', and various 'GMO' and 'FMO' identifiers. The main area displays a grid of microscopy images. On the right, a metadata panel for image 'FMO02796\_conf\_130829\_2013\_08\_29\_11\_' is visible, showing details such as 'IMAGE ID: 4843', 'Owner: Aris Polyzos', 'Acquisition Date: 2013-08-29 17:17:44', 'Imported Date: 2013-08-29 21:08:46', 'Dimensions (XY): 512 x 512', 'Pixels Type: uint16', 'Pixels Size (XYZ) (µm): 0.20 x 0.20 x 1.00', 'Z-sections/Timepoints: 15 x 1', and 'Channels: DAPI, Anti-HA, phalloidin'. There are also sections for 'RATING' and 'TAGS'.

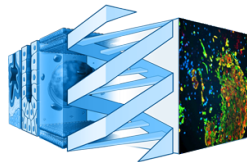


# Image acquisition workflow

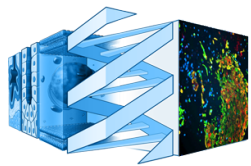
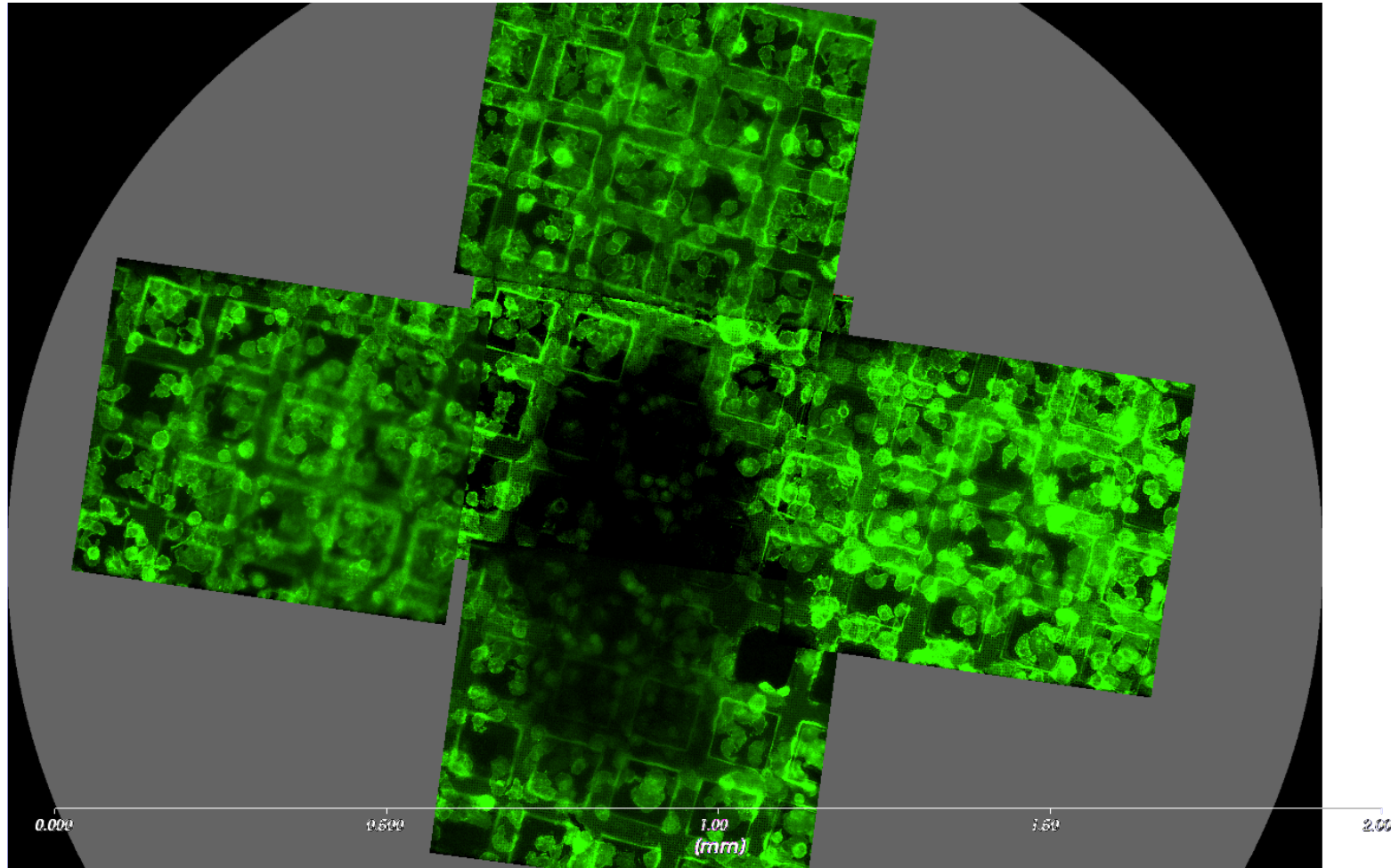


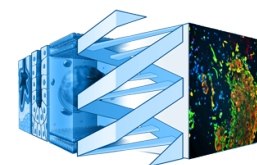
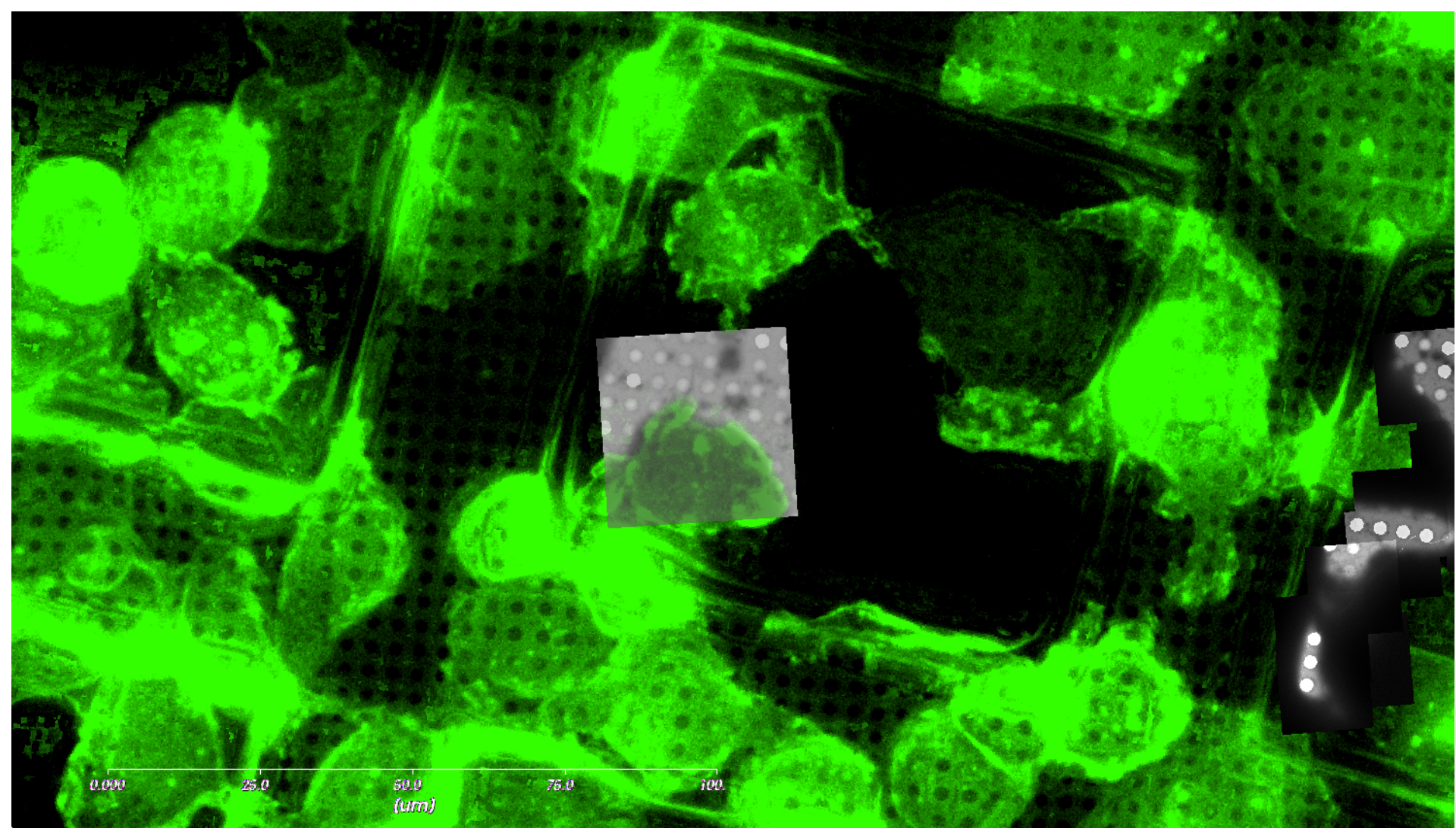
# Our experience

- Likes:
  - Easy setup (after dealing with OHSU firewall issues)
  - Excellent retrieval performance: inside/outside OHSU
  - Fully user-initiated image uploads and image retrieval
  - Good support for our image/experiment types
- Wishes:
  - Support for rich experimental metadata (e.g. ISA-Tab)
  - Bulk image download
  - Import using OMERO.web
  - Deeper folder structure

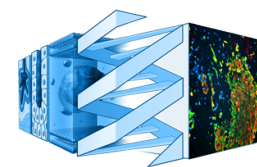
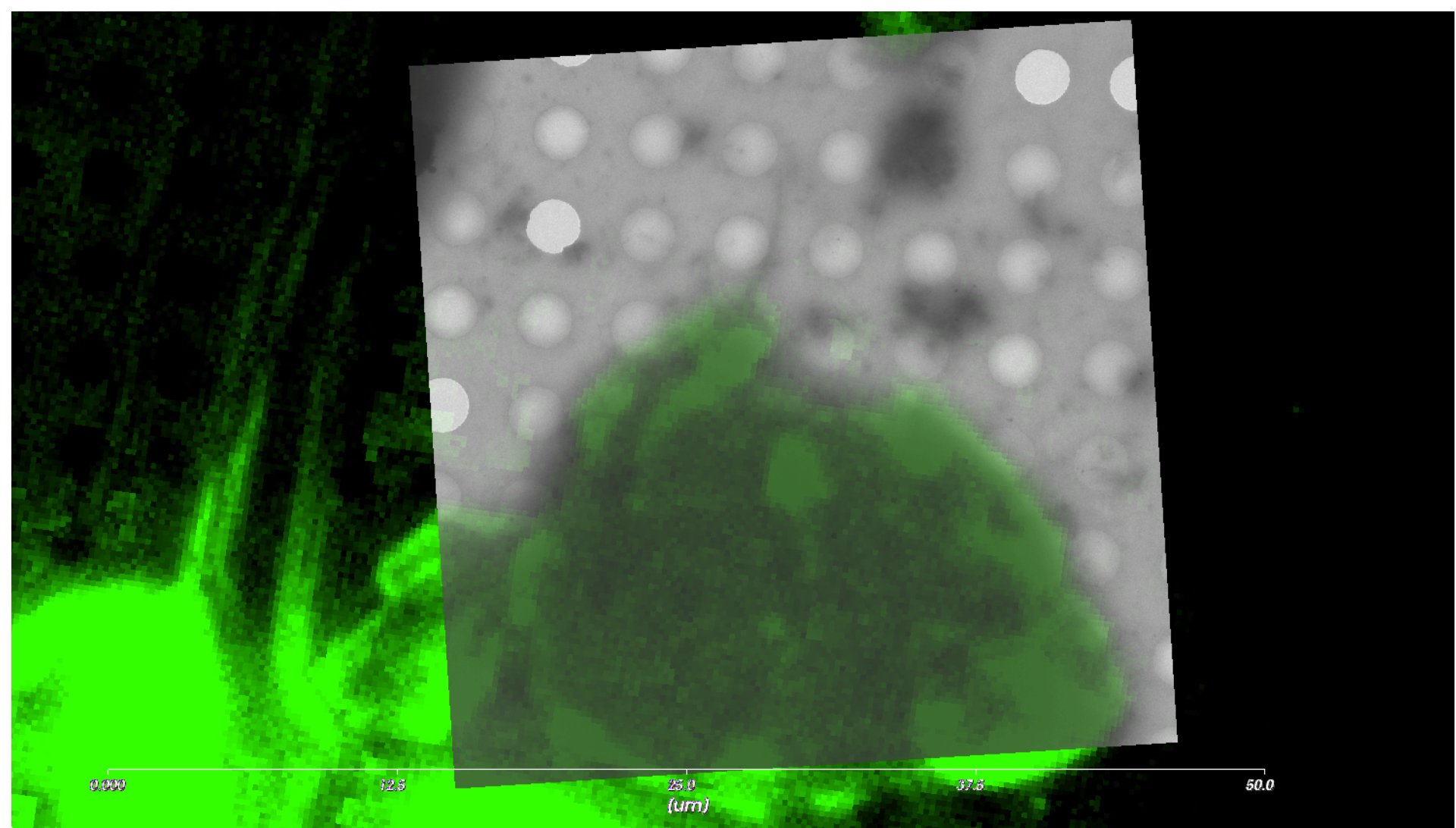


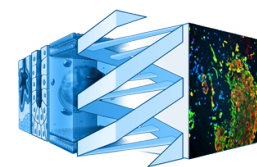
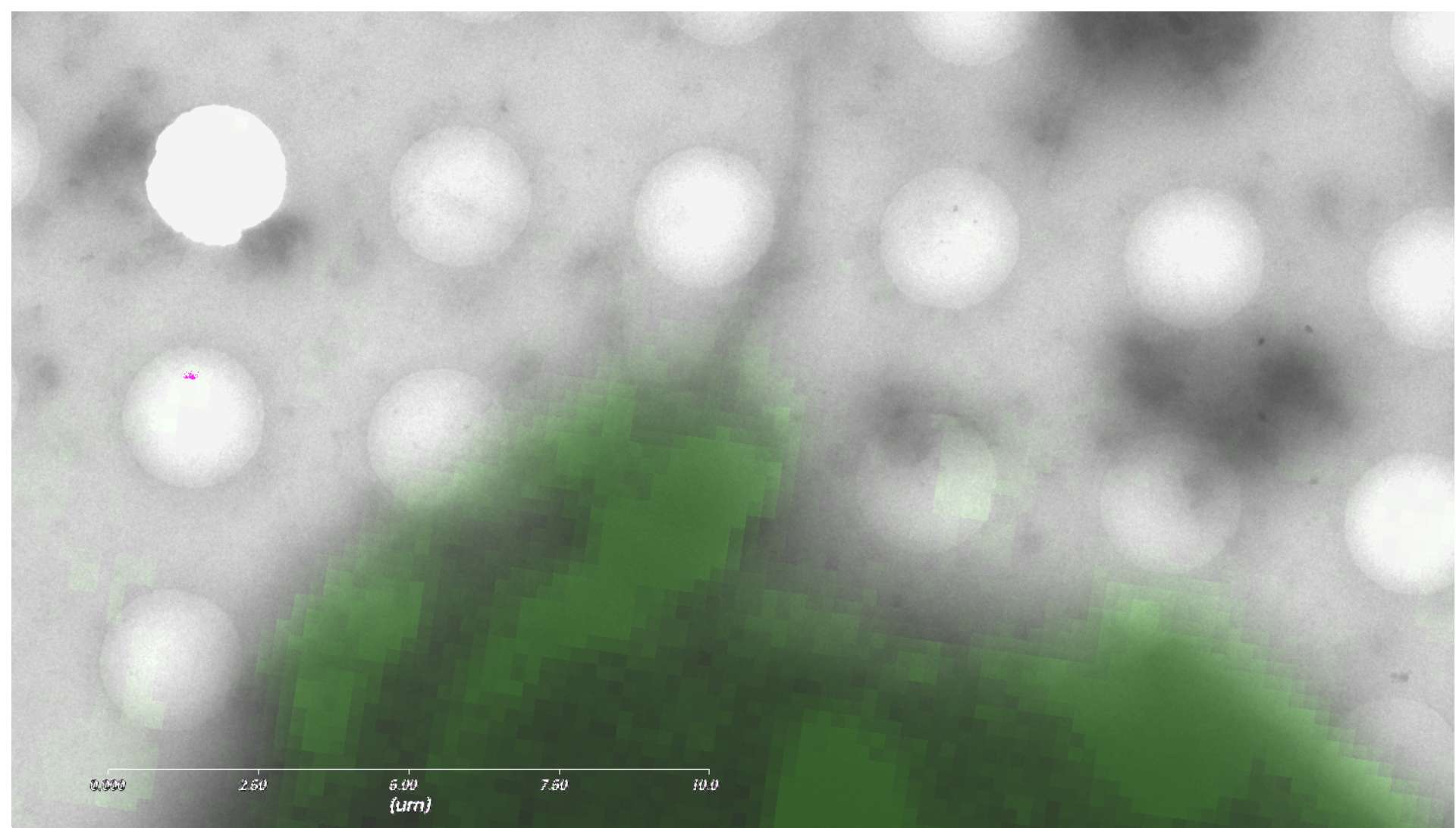
# 2D iLEM workflow w/ iCorr

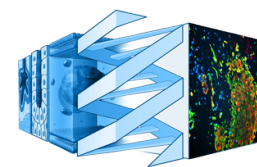
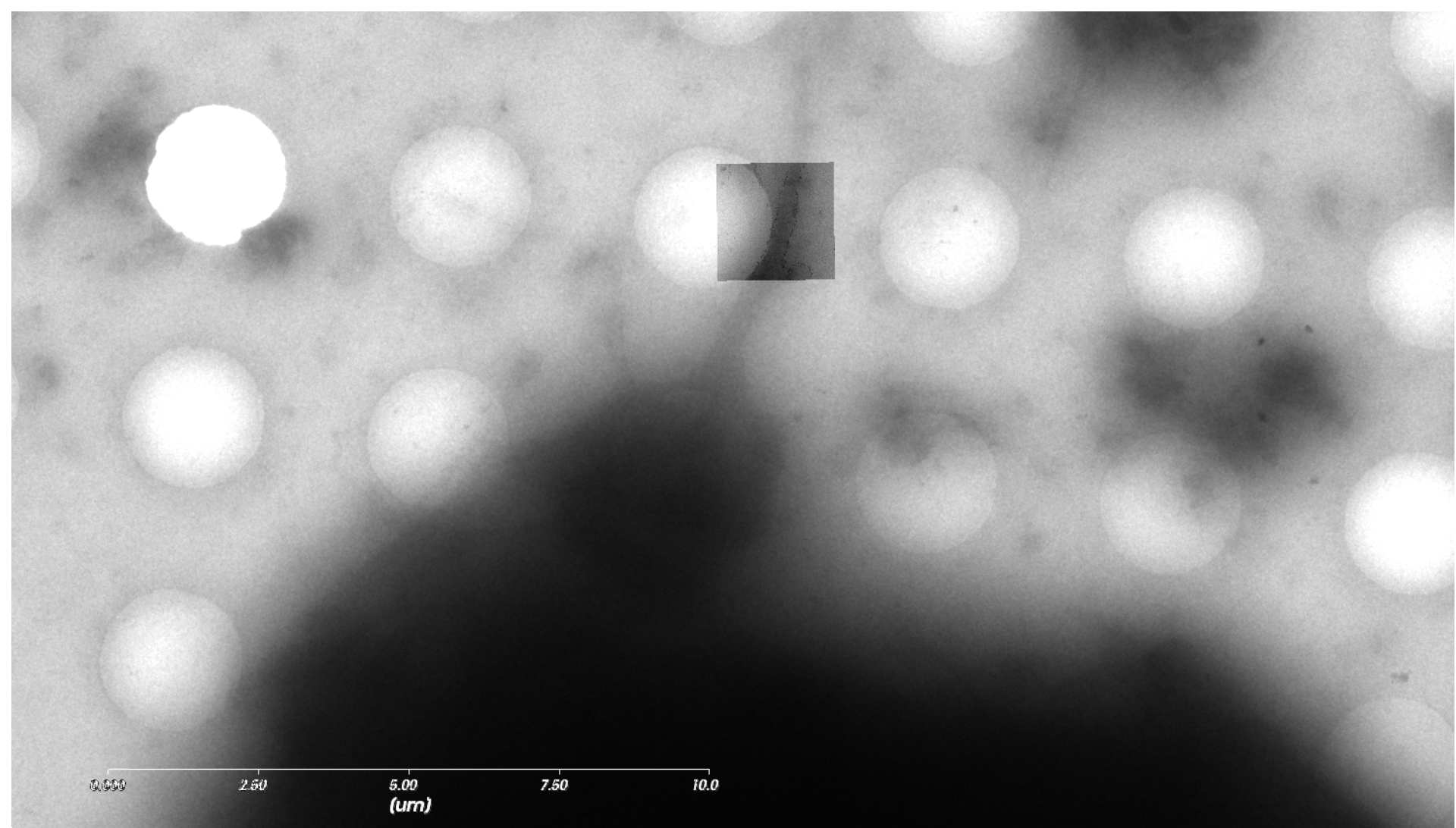


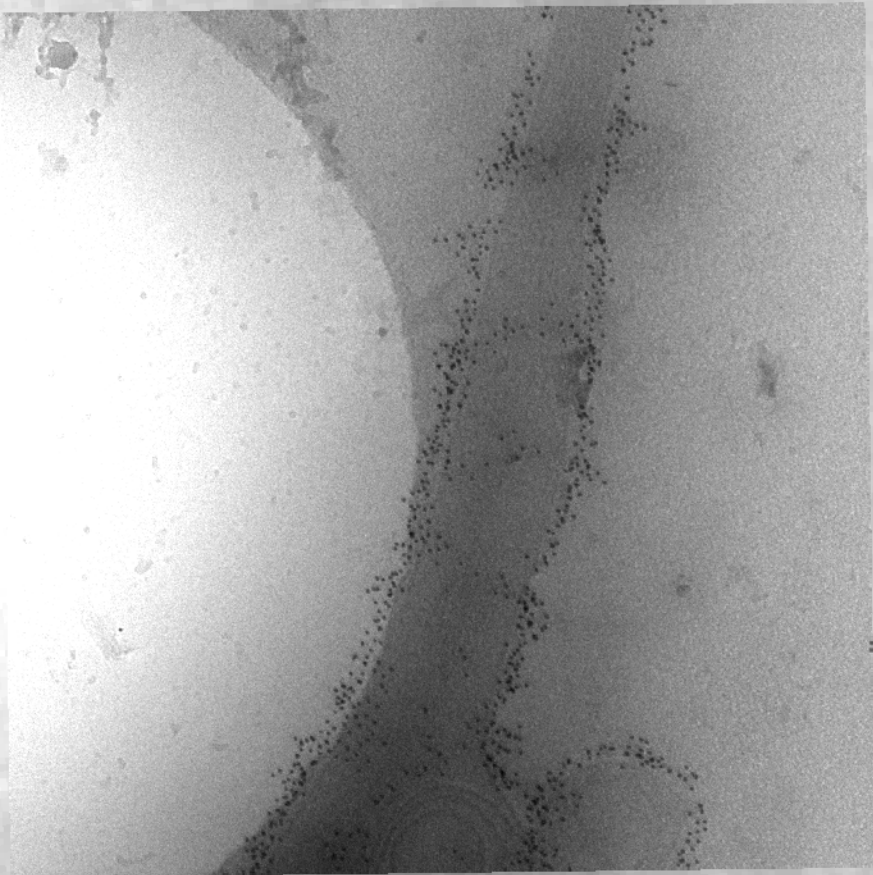












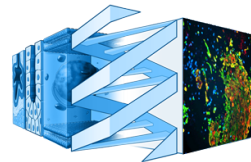
0.000

0.750

1.50  
( $\mu\text{m}$ )

2.25

3.00



# 3D iLEM workflow w/ FIB/SEM

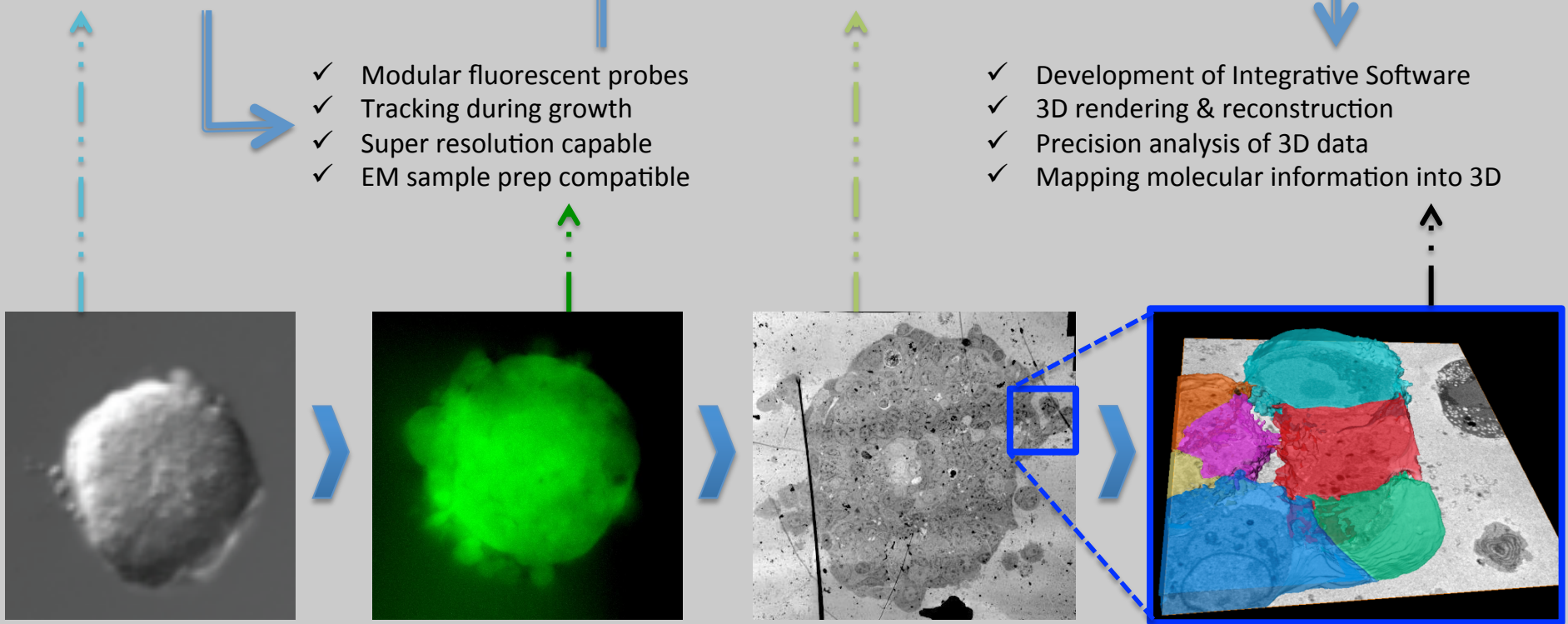
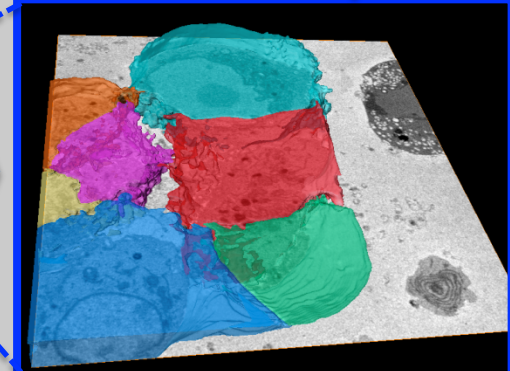
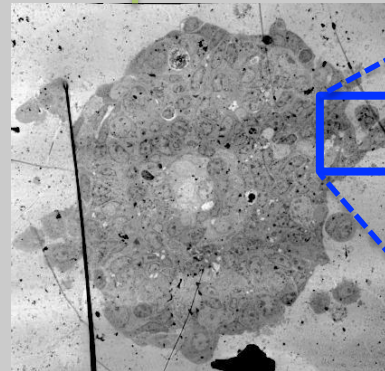
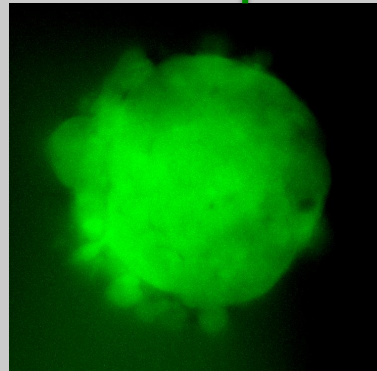
## Integrating molecular information and structural detail with nanometer precision

- ✓ Multicellular growth (Organovo)
- ✓ Printing wide spectrum of tissues
- ✓ Disease oriented research
- ✓ Live cell imaging

- ✓ Cryo-immobilization (HPF & plunge)
- ✓ Preservation of probes in EM prep
- ✓ Multiple EMs (Krios, Helios, iCorr)
- ✓ MAPS correlation of SEM data

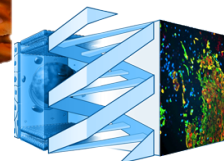
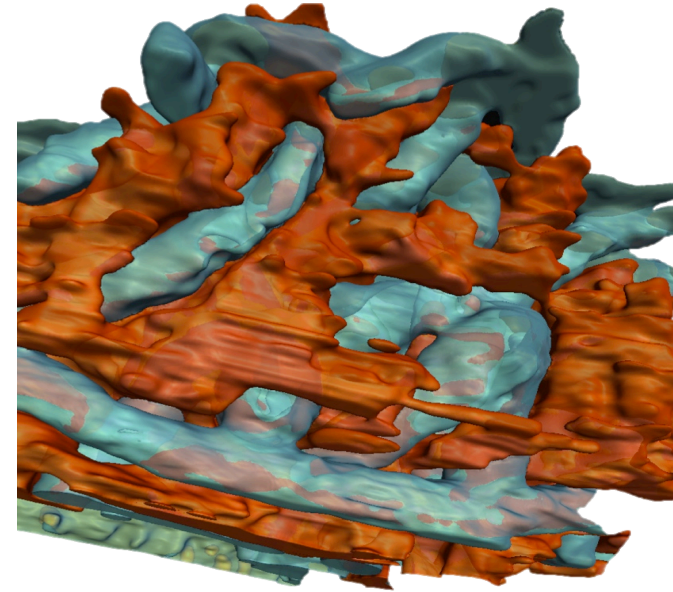
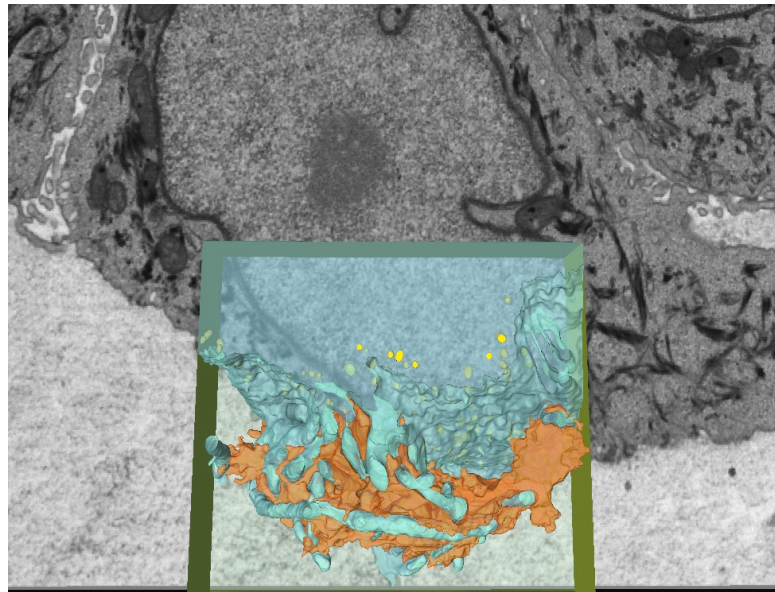
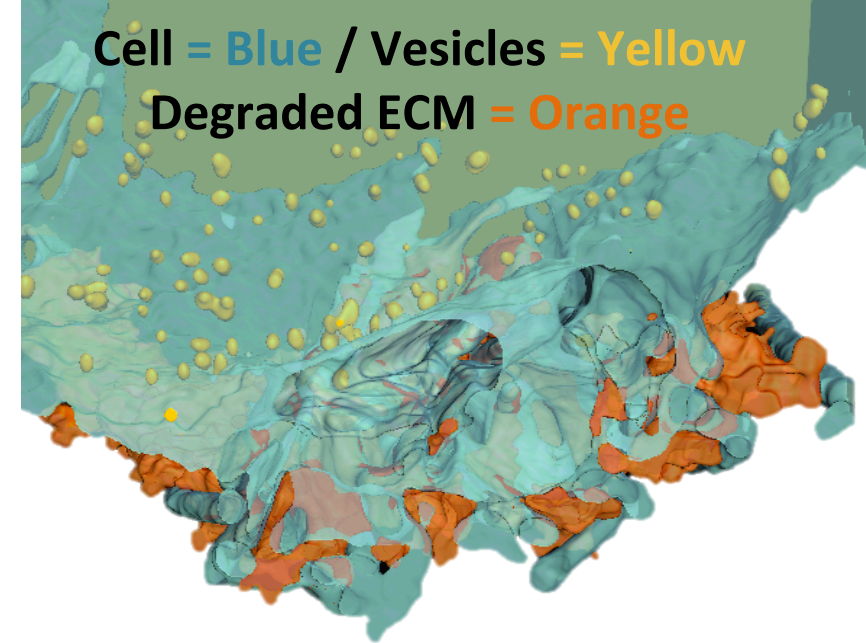
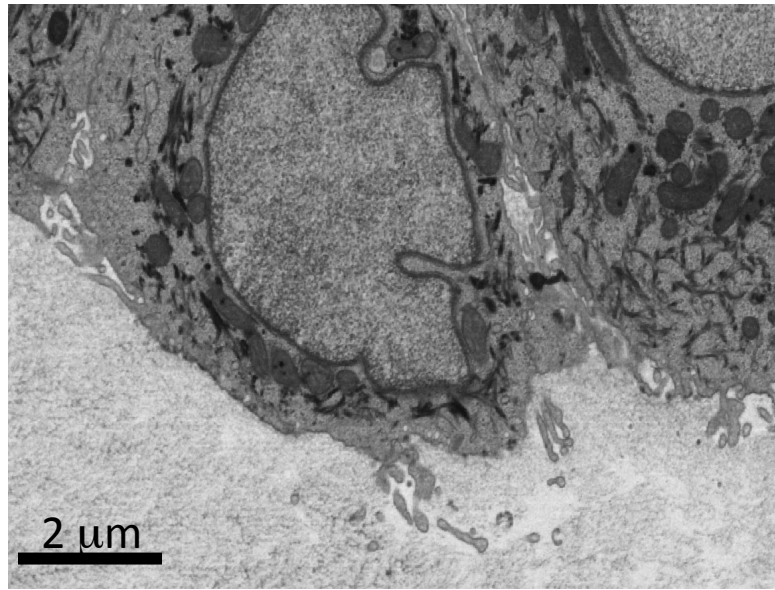
- ✓ Modular fluorescent probes
- ✓ Tracking during growth
- ✓ Super resolution capable
- ✓ EM sample prep compatible

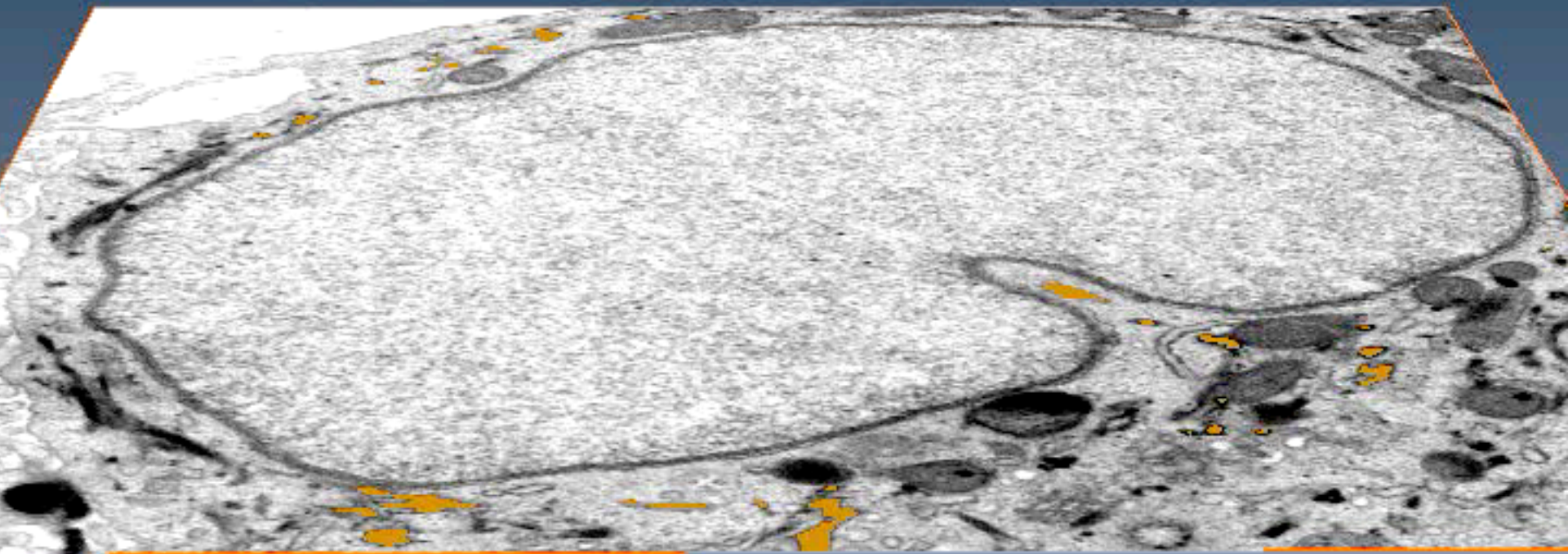
- ✓ Development of Integrative Software
- ✓ 3D rendering & reconstruction
- ✓ Precision analysis of 3D data
- ✓ Mapping molecular information into 3D



# Integrating molecular information and structural detail with nanometer precision

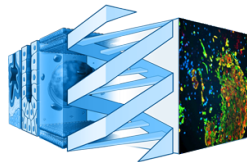
Helios Nanolab 650  
FIB-SEM





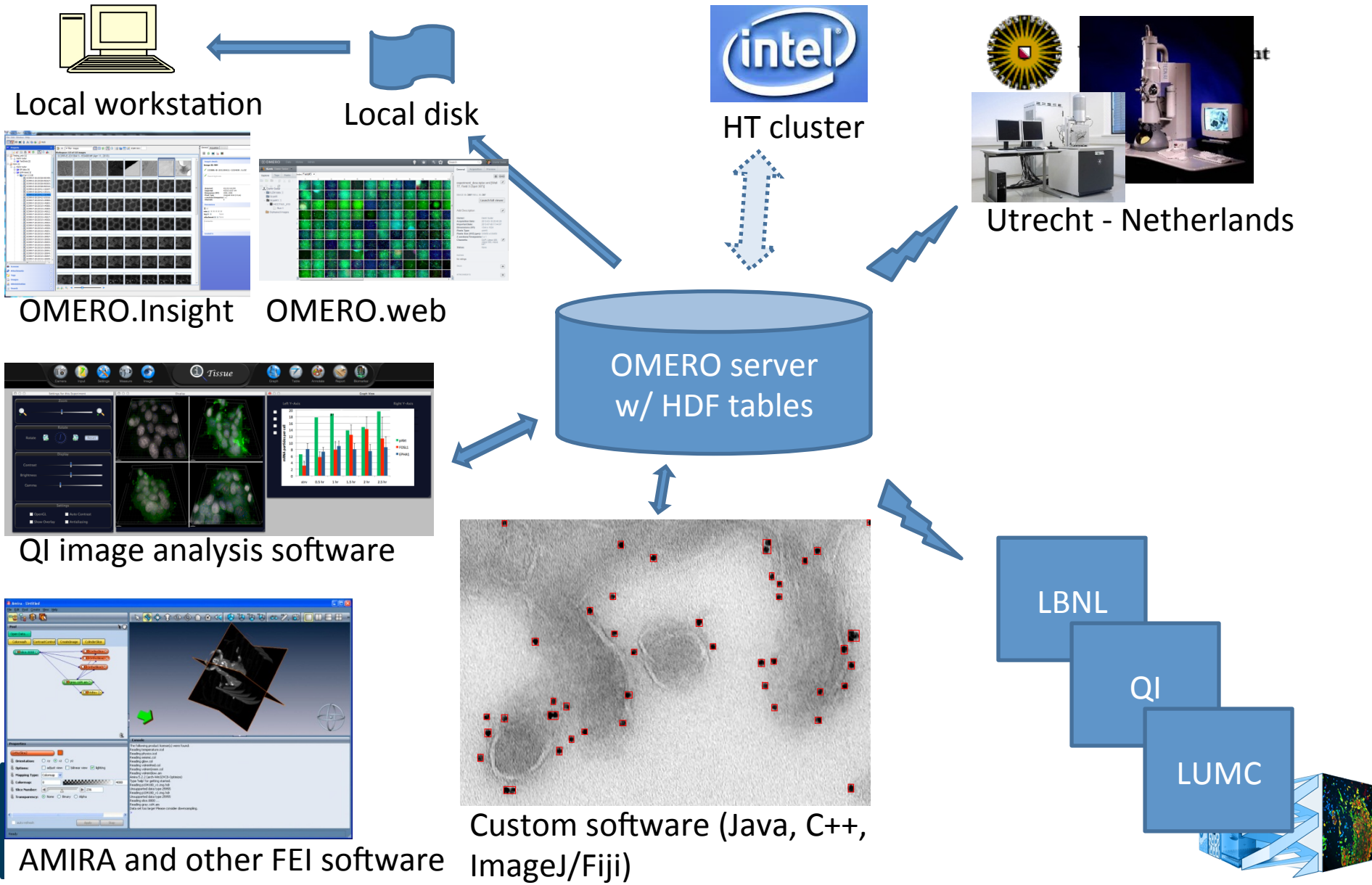
# Our experience

- Likes:
  - Large images are handled well
  - Scripting
  - Flexible data access options
- Wishes:
  - Registration information between imaging modalities i.e. a Real-World coordinate system
  - Tiled/chunked storage of nD data in multiple scales
  - Automated segmentation methods



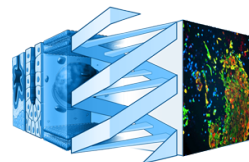
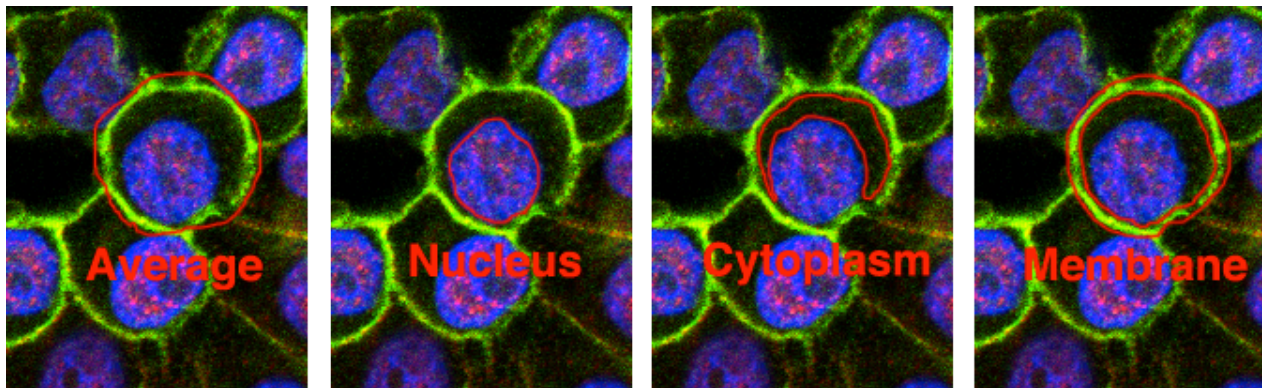


# Image analysis/visualization workflow



# Image analysis and visualization

- Find individual cells
- Find sub cellular regions: Nucleus, Cytoplasm, Membrane
- Measure biomarker expressions in each region
- Relate channels: e.g. pAkt versus total Akt
- Show statistics & relate statistics to morphology of cells and tissue
- Compare over time and perturbation conditions





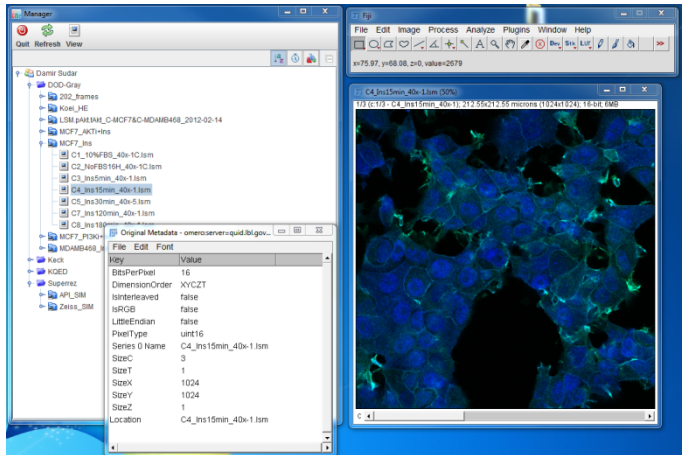
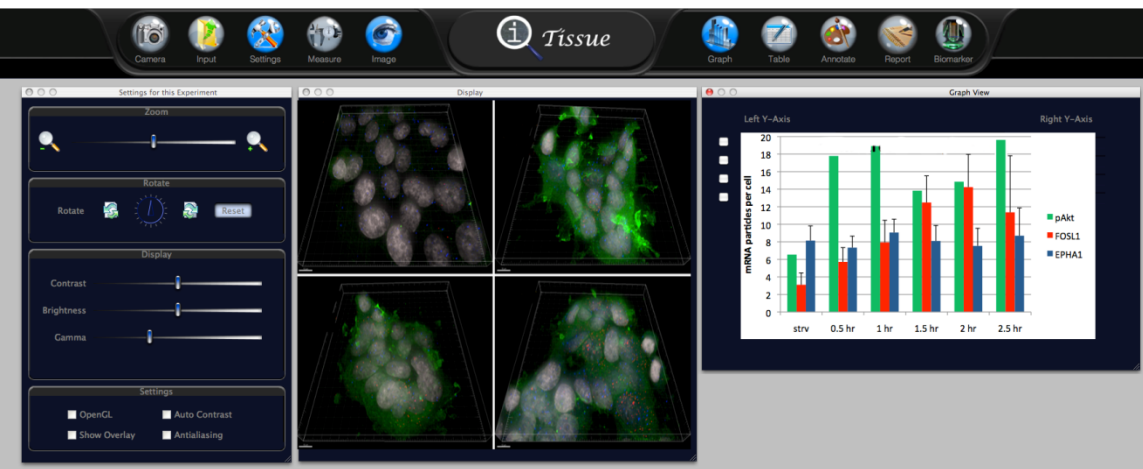
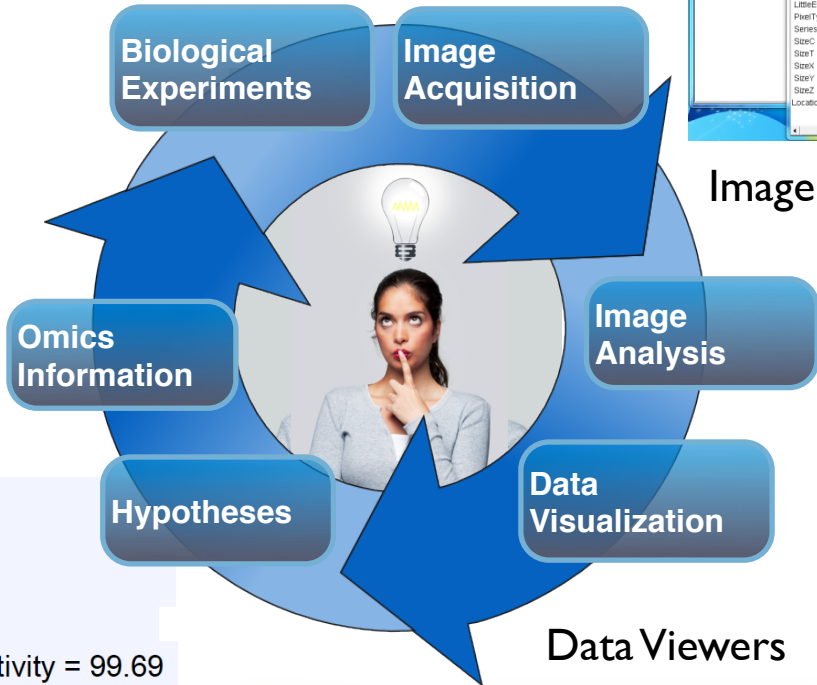
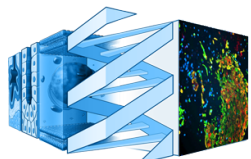
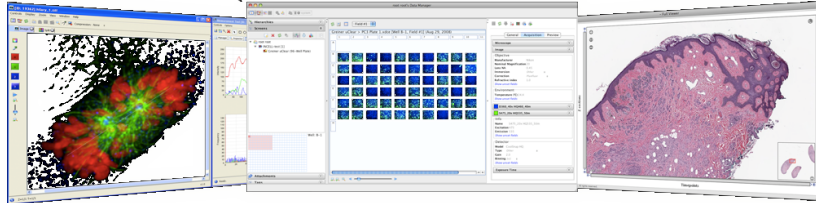


Image & Metadata Database

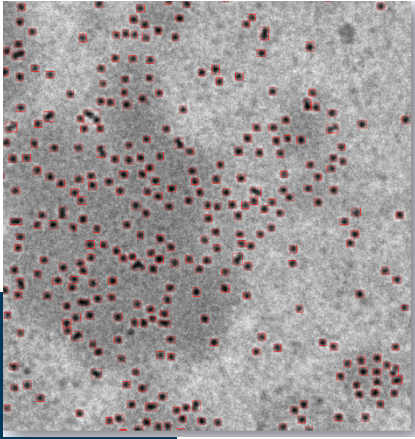
Qi Fluorescence Biomarker Quantitation



Data Viewers



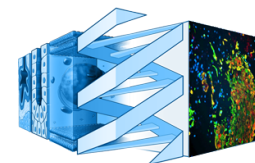
EM Image Analysis



Sensitivity = 99.69  
Specificity = 98.66

# Our experience

- Likes:
  - Good Python support
  - Access to original data via “Archived” or “File paths”
  - HDF support for feature data
- Wishes:
  - Improved Java and C++ API support and documentation
  - Well defined infrastructure to initiate server-side processing on HPC cluster
  - Easy use/display/searching of the HDF data
  - Abstracted shape descriptions of objects in close relationship with pixel/voxel descriptions



# Acknowledging



Manfred Auer, Walter Georgescu,  
Aris Polyzos, David Skinner, Joaquin  
Correa, Erwin Frise



Joe Gray, Danielle Jorgens, Anke  
Mulder, Guillaume Thibault, Michel  
Nederlof (QI), Jan Post (UU), Elly van  
Donselaar (UU)

And the OME team!!!

